

Systematic Investigation into the Matsuda-Heck Reaction of lpha-Methylene Lactones: How Conformational Constraints Direct the β -H-Elimination Step

Bernd Schmidt,* Felix Wolf, and Christopher Ehlert

Universitaet Potsdam, Institut fuer Chemie, Karl-Liebknecht-Straße 24-25, D-14476 Potsdam-Golm, Germany

Supporting Information

ABSTRACT: α -Methylene- γ -butyrolactone and α -methyleneδ-valerolactone undergo Pd-catalyzed Matsuda-Heck couplings with arene diazonium salts to α -benzyl butenolides or pentenolides, respectively, or to α -benzylidene lactones. The observed regioselectivity is strongly ring size dependent, with six-membered rings giving exclusively α -benzyl pentenolides, whereas the five-membered α -methylene lactone reacts to mixtures of regioisomers with a high proportion of (E)- α benzylidene-γ-butyrolactones. DFT calculations suggest that the reasons for these differences are not thermodynamic but kinetic in nature. The relative energies of the conformers of

endo-
$$\beta$$
-H elimination

Ar-N₂ BF₄
Pd(0)

By Helimination

moderate to low selectivity

the Pd σ -complexes resulting from insertion into the Pd-aryl bond were correlated with the dihedral angles between Pd and endo- β -H. This correlation revealed that in the case of the six-membered lactone an energetically favorable conformer adopts a nearly synperiplanar Pd/endo- β -H arrangement, whereas for the analogous Pd σ -complex of the five-membered lactone the smallest Pd/endo-β-H dihedral angle is observed for a conformer with a comparatively high potential energy. The optimized conditions for Matsuda-Heck arylations of exo-methylene lactones were eventually applied to the synthesis of the natural product anemarcoumarin A.

INTRODUCTION

Acrylates 1 are highly reactive benchmark olefins for evaluating novel catalyst systems or reaction conditions for Mizoroki-Heck couplings. The high π -acceptor and poor σ -donor reactivity of acrylates leads to an electronically biased insertion of the olefin into the Pd- σ -aryl bond of 2, which results in the formation of a Pd σ -complex 3 with the new C-C-bond in the β -position and a Pd-C^α σ-bond. The catalytic cycle proceeds with a β -hydride elimination, which requires a *syn*-periplanar or nearly syn-periplanar arrangement of the Pd at C^{α} and one β hydrogen. To adopt the energetically least unfavorable synperiplanar conformation 3', a rotation around the $C^{\beta}-C^{\alpha}$ bond must occur. The stereospecificity of the β -H-elimination and the preferred anti-orientation of the aryl and ester substituents explain the very high E-selectivity observed for the cinnamates 5 (Scheme 1a). While a plethora of examples for Mizoroki-Heck arylations of simple acrylates exist, comparatively little is known about the behavior of conformationally constrained enoates in this transformation. One type of conformationally restricted enoates are α,β -unsaturated lactones 6. If the generally accepted mechanism for Heck-type arylations as outlined in Scheme 1a is applied to these substrates, a Pd σ complex 7 results, which cannot undergo the syn-β-Helimination required to close the catalytic cycle. Nevertheless some examples for successful Mizoroki-Heck reactions of cycloalkenes, including α,β -unsaturated six-membered lactones,⁵ lactams,^{5,6} and cycloketones,^{7,8} have been described in

the literature. For lack of a plausible $syn-\beta$ -H elimination pathway, base-induced *trans-\beta-H-elimination*⁴ or epimerization⁷ of the Pd σ-complex 7 to epi-7 via a Pd enolate 7' have been proposed to explain the formation of arylated products 8 (Scheme 1b).

Another, even less investigated type of conformationally restricted enoates are exo-methylene lactones 9. The Pd σ complex **10** resulting from insertion into the Pd $-\sigma$ -aryl bond of **2** can in principle undergo β -H-elimination with one $exo-\beta$ -H and the *syn-endo-\beta-H*, leading to α -benzylated- α , β -unsaturated lactones endo-11 or α -benzylidene lactones exo-11 (Scheme 1c). For example, Genêt and co-workers reported that the Pdcatalyzed coupling of an arene diazonium salt with α methylene-γ-butyrolactone (9a) gives exclusively the exobenzylidene lactone (exo-11) as a mixture of E- and Z-isomers. Shortly afterward, Arcadi et al. investigated the coupling of 9a with various aryl iodides and found that mixtures of endo- and *exo*-products were formed and that the *exo*-products are exclusively Z-configured. For more densely substituted α methylene-γ-butyrolactones, Kim et al. found that both via Mizoroki-Heck arylation or via oxidative C-H-activation mixtures of endo- and E-exo-arylation products were formed. 12 The exclusive formation of exo-arylation products as E/Zmixtures was reported for guaianolide-type sesquiterpene

Received: September 8, 2016 Published: October 17, 2016

Scheme 1. Heck-Type Arylations of Acrylates, Unsaturated Lactones, and α -Methylene Lactones

a) Mizoroki-Heck arylation of acrylates

b) Mizoroki-Heck arylation of α,β-unsaturated lactones

c) Mizoroki-Heck arylation of α -methylene lactones

lactones. ¹³ To date, the above-mentioned studies by Arcadi et al. remain the only systematic investigations into Mizoroki—Heck reactions of α -methylene lactones. ^{10,11} From their observations, these authors concluded that the electronic effects of *para*-substituents at the aryl iodide and the nature of the base employed in the coupling reaction steer the selectivity. Thus, particularly high *endo*-selectivities were observed for aryl iodides with an electron-withdrawing *para*-substituent and with acetates rather than amines as a base. As a rationale for the beneficial effect of acetate, the authors proposed that Pd-bound acetate participates in the β -H-abstraction through a seven-membered transition state. Over the past few years, we ^{14–17} and others ^{18–23} reported a

Over the past few years, we^{14–17} and others^{18–23} reported a considerable number of examples for Heck-type coupling reactions with arene diazonium salts. This variant of the Mizoroki–Heck coupling, often referred to as the Matsuda–Heck reaction, was originally discovered in the late 1970s by Matsuda and co-workers but lay dormant for many years. ^{24,25} Over the past 15 years, Matsuda–Heck- and other Pd-catalyzed cross couplings of arene diazonium salts have attracted renewed and still growing interest due to a number of beneficial features. ^{26–28} For instance, diazonium salts are conveniently synthesized from amines or acetamides²⁹ (or can be generated in situ, ³⁰ e.g., under flow conditions ³¹), are highly reactive,

allow for monitoring the kinetics by quantifying the nitrogen evolution over time, 32 and do not require elaborate ligands for catalyst tuning (Pd(OAc)₂ and Pd₂(dba)₃ are the most commonly used precatalysts). Finally, Matsuda-Heck reactions can be conducted both under basic and base-free conditions, 32,33 which, in light of the assumed vital role of acetate bases in Arcadi's studies, prompted us to revisit Heck-type arylations of exo-methylene lactone 9a using arene diazonium salts. Surprisingly, the homologous six-membered exo-methylene lactone 9b or substituted derivatives of the same ring size have, to the best of our knowledge, never been investigated in Heck-type reactions. As we expected that the ring size affects the $Pd-C^{\alpha}-C^{\beta endo}-H$ dihedral angle and hence the regioselectivity of the β -H-elimination step, we included 9b, its δ methyl derivative **9c**, and the *N*-benzyl δ -valerolactam **9d** in our study.

RESULTS AND DISCUSSION

Syntheses of *exo*-Methylene Carbonyl Compounds 9 and Diazonium Salt 13m. *exo*-Methylene γ -butyrolactone (9a) is widely commercially available from a number of suppliers. It has, like its less conveniently accessible sixmembered homologue 9b, been synthesized from the corresponding lactone by deprotonation, Claisen condensation with ethyl formate, and addition of formaldehyde. Several 2-methylene-δ-valerolactones, including 9c, have been synthesized by conjugate addition of carbonyl compounds to α-phosphono acrylates, subsequent lactonization, and olefination with formaldehyde. For the synthesis of 9b and 9c, we used a modification of the former method which was adapted from a synthesis of malyngolide but had not been applied to these particular derivatives (Scheme 2).

Scheme 2. Syntheses of α -Methylene- δ -valerolactones 9b,c

N-Benzyl- α -methylene- δ -valerolactam (9d) was recently synthesized from δ -valerolactam in three steps by aldol condensation with formaldehyde, but we decided to use a two-step synthesis starting from ethyl nipecotate (12d). This compound was N-benzylated to N-Bn-12d and then subjected to a base-induced α -methylenelactam rearrangement ^{39,40} to furnish 9d (Scheme 3).

These syntheses can be routinely performed on a 20 mmol scale and reliably provide gram quantities of the required *exo*methylene lactones and lactams **9**.

Scheme 3. Synthesis of α -Methylene- δ -valerolactam (9d)

The arene diazonium salts used in this study were obtained either by diazotation of anilines or via deacetylation—diazotation of acetanilides following previously published procedures. The hitherto unknown phenol diazonium salt 13l was synthesized from methyl 4-hydroxyanthranilate (14l) through diazotation with NaNO₂ in aqueous HBF₄ (Scheme 4).

Scheme 4. Synthesis of the Hitherto Unknown Diazonium Salt 13l

Matsuda Heck Arylation of α -Methylene- δ -valerolactone (9b). For the purpose of optimization, we investigated the Matsuda-Heck coupling of 9b and 4-methoxybenzene diazonium salt 13a. Although other solvents, in particular, water, 43-45 have been tested for Heck-type coupling reactions of arene diazonium salts, methanol 32,41,46 and acetonitrile, 47,48 either under basic or under base-free conditions, are still the most commonly used solvents (Table 1). Base-free conditions in methanol (entries 1 and 2) were chosen for comparison of the catalysts. Pd(OAc), was found to perform substantially better than Pd2dba3·CHCl3 and was, therefore, used in all other experiments. By addition of NaOAc as a base (entry 3), the yield of the Matsuda-Heck coupling product could be further increased to 80%. In marked contrast, the reaction fails completely in acetonitrile in the absence of NaOAc (entry 4). This observation is in line with previous findings by us^{14,32} and Correia's mechanistic studies, 47 which suggest that acetonitrile serves as a stabilizing ligand for the Pd hydride species resulting from the β -hydride elimination step. Consequently, added base should facilitate the regeneration of the catalytically active Pd(0) species. Upon addition of NaOAc, we obtained indeed a nearly quantitative yield of the coupling product (entry 5).

We then investigated a reduced catalyst loading (entry 6), which leads to a dramatically diminished yield. The same was observed when the excess of alkene **9b** was reduced (entry 7). Using the diazonium salt **13a** with an excess of 1 equiv (entry 9) led to an improved yield compared to a 10% excess (entry 8), but application of the *exo*-methylene lactone **9b** in excess appears to be more suitable and more convenient to ensure high yields of the desired coupling product. Careful examination of the crude reaction mixtures revealed that in all cases *endo*-**11ba** was exclusively formed.

Scope and limitations of Matsuda-Heck arylations of 9b were explored for a number of other arene diazonium salts 13 using 5 mol % of Pd(OAc)₂ in all experiments and a ratio of reactants of 2:1 (9b/13). As solvents, either methanol or acetonitrile (with and without added base) was tested for each diazonium salt. For most diazonium salts, at least one set of conditions was identified to obtain the respective Matsuda-Heck products endo-11b# in synthetically useful yields and without the formation of any exo-isomers (Table 2). These results show that methanol without added base is usually the optimal condition and that acetonitrile with added base is only in exceptional cases superior. This includes the example chosen for the optimization study, which underlines once again that the optimum reaction conditions of Matsuda-Heck reactions are strongly substrate dependent and that generalizations must be made with care.

A perspective application of these Matsuda—Heck products in stereoselective synthesis is their conversion to 2-substituted 2,4-dienoic acids through base-induced elimination. This transformation was originally discovered as early as 1859 for the conversion of the rowan berry oil constituent parasorbic acid to sorbic acid. More recently, the reaction was applied to the total synthesis of complex natural products with conjugated diene moieties, $^{50-53}$ and very recently, we demonstrated that a base-mediated eliminative ring opening of β , γ -unsaturated lactones can be incorporated in a tethered ring-closing metathesis sequence. $^{54-56}$

We are not aware of examples for the eliminative ring opening of α -benzylated pentenolides and therefore inves-

Table 1. Optimization of Matsuda-Heck Conditions for 9b

entry	ratio 9b:13a	precatalyst (catalyst loading)	base ^a	solvent	product (yield, %) ^b
1	2.0:1.0	Pd ₂ (dba) ₃ ·CHCl ₃ (2.5 mol %)		CH ₃ OH	endo-11ba (54)
2	2.0:1.0	Pd(OAc) ₂ (5.0 mol %)		CH ₃ OH	endo- 11ba (64)
3	2.0:1.0	Pd(OAc) ₂ (5.0 mol %)	NaOAc	CH ₃ OH	endo- 11ba (80)
4	2.0:1.0	Pd(OAc) ₂ (5.0 mol %)		CH ₃ CN	с
5	2.0:1.0	Pd(OAc) ₂ (5.0 mol %)	NaOAc	CH ₃ CN	endo- 11ba (98)
6	2.0:1.0	Pd(OAc) ₂ (2.5 mol %)	NaOAc	CH ₃ CN	endo- 11ba (61)
7	1.2:1.0	Pd(OAc) ₂ (5.0 mol %)	NaOAc	CH ₃ CN	endo- 11ba (50)
8	1.0:1.1	Pd(OAc) ₂ (5.0 mol %)	NaOAc	CH ₃ CN	endo- 11ba (38)
9	1.0:2.0	Pd(OAc) ₂ (5.0 mol %)	NaOAc	CH ₃ CN	endo- 11ba (71)

^a3.0 equiv. ^bOnly *endo-***11ba** was observed in all experiments. ^cNo conversion (TLC).

Table 2. Scope and Limitations of Matsuda-Heck Couplings with 9b

entry	13	\mathbb{R}^1	\mathbb{R}^2	\mathbb{R}^3	solvent	base	product	yield (%)
1	13a	Н	Н	OMe	CH ₃ CN	NaOAc	endo-11ba	98
2	13b	Н	Н	ОН	CH ₃ OH		endo-11bb	74
3	13c	Н	Н	OBn	CH ₃ CN	NaOAc	endo-11bc	66
4	13d	Н	Br	OMe	CH ₃ OH		endo-11bd	82
5	13e	Н	Br	ОН	CH ₃ OH	NaOAc	endo-11be	68
6	13f	Н	NO_2	OMe	CH ₃ OH		endo-11bf	82
7	13g	Н	NO_2	ОН	CH ₃ OH		endo-11bg	99
8	13h	NO_2	Н	OMe	CH ₃ OH		endo-11bh	48
9	13i	Н	CF ₃	Н	CH ₃ OH		endo-11bi	а
10	13j	Н	Н	NO_2	CH ₃ OH		endo-11bj	67
11	13k	Н	CO ₂ Me	ОН	CH ₃ OH		endo-11bk	а
12	131	CO_2Me	Н	ОН	CH ₃ OH		endo-11bl	49
13	13m	Н	Н	NHAc	CH ₃ CN	NaOAc	endo-11bm	31

^aNo conversion in methanol or acetonitrile under either basic or base-free conditions.

tigated the base-induced cleavage of *endo-11ba*. Treatment of this lactone with KOH in DMSO at slightly elevated temperature resulted in the clean formation of the 2-benzylated 2,4-dienoic acid *Z-15ba* within 1 h. Assignment of the *Z*-configuration as shown in Scheme 5 is based on an NOE

Scheme 5. Base-Induced Eliminative Ring Opening of α -Benzyl Pentenolides

interaction between H^3 and the CH_2 group of the benzyl substituent. To check whether a stereoselective formation of the C^4-C^5 double bond is also possible, we synthesized the pentenolide *endo-11ca* from α -methylene lactone 9c under the same conditions used for *endo-11ba* (see Table 1, entry 5) and subjected this compound to the base-induced eliminative ring opening. The conjugated diene (2Z,4E)-15ca was isolated in a comparable yield as a single isomer. Assignment of the 2Z-configuration was again achieved by NOE interactions between H^3 and the benzylic CH_2 group, and the 4E-configuration was proved by an NOE interaction between the terminal methyl group and H^4 as well as a $^3J(H^4,H^5)$ value of 15.0 Hz (Scheme 5).

Matsuda Heck Arylation of N-Benzyl α -Methylene- δ -valerolactam (9d). When we applied the optimized

conditions for the Matsuda-Heck coupling of phenoldiazonium salt 13b and α -methylene lactone 9b (base-free methanol, 2.0:1.0 ratio of alkene and diazonium salt) to the coupling of this diazonium salt and α -methylene lactam 9d, we could isolate endo-11db in 61% yield. As for the analogous lactones, no exoisomers could be detected in the reaction mixture. In contrast to the results observed during the optimization study for Matsuda-Heck couplings with lactone 9b, we noted an increased yield of 75% when the ratio of 9d to 13b was reduced to 1.2:1.0. After testing some other ratios of reactants, we eventually found that the highest yield was obtained for a 2.0:1.0 ratio of diazonium salt 13 and exo-methylene lactam 9d. Scope and limitations of Matsuda–Heck arylations of 9d with various arene diazonium salts 13 were next evaluated, routinely using a 1.0:2.0 ratio of reactants. Both methanol and acetonitrile were tested with and without added base for each diazonium salt. Apart from base-free acetonitrile, which failed to give a substantial yield of coupling products in all cases, most diazonium salts could be coupled with 9d in synthetically useful yields under at least one set of conditions (Table 3). Exceptions are the *m*-trifluoromethyl-substituted arene diazonium salt 13i, which failed to undergo Matsuda-Heck couplings with both lactone 9b and lactam 9d (entry 9) and diazonium salts 13h and 13m. Both underwent coupling reactions with 9b in moderate yields, but not with 9d (entries 8 and 13). On the other hand, methyl carboxylate substituted diazonium salt 13k, which did not react with lactone 9b, reacted in a satisfying yield with the lactam (entry 11). In general, the isolated yields of Matsuda-Heck coupling products are lower for the lactam 9d compared to the analogous lactone 9b. Although the arene diazonium salts were used in excess, we did not notice the formation of symmetrical biaryls. This transformation has very recently been described when diazonium salts were exposed to catalytic amounts of Pd(OAc)₂ in ionic liquids.⁵

Matsuda Heck Arylation of α -Methylene- γ -butyrolactone (9a). In contrast to the Matsuda-Heck couplings

Table 3. Scope and Limitations of Matsuda-Heck Couplings with 9d

13	(2.0)	equiv.)
	(2.0	cquiv.

entry	13	\mathbb{R}^1	\mathbb{R}^2	\mathbb{R}^3	solvent	base	product	yield (%)	
1	13a	Н	Н	OMe	CH ₃ CN	NaOAc	endo-11da	72	
2	13b	Н	Н	ОН	CH ₃ OH		endo-11db	86	
3	13c	Н	Н	OBn	CH ₃ CN	NaOAc	endo-11dc	65	
4	13d	Н	Br	OMe	CH ₃ OH		endo-11dd	76	
5	13n	Н	Br	OBn	CH ₃ OH		endo-11dn	66	
6	13f	Н	NO_2	OMe	CH ₃ OH		endo-11df	80	
7	13g	Н	NO_2	ОН	CH ₃ OH		endo-11dg	96	
8	13h	NO_2	Н	OMe	CH ₃ OH		endo-11dh	а	
9	13i	Н	CF ₃	Н	CH ₃ OH		endo-11di	а	
10	13j	Н	Н	NO_2	CH ₃ OH		endo-11 d j	34	
11	13k	Н	CO ₂ Me	ОН	CH ₃ OH		endo-11dk	66	
12	131	CO ₂ Me	Н	ОН	CH ₃ OH		endo-11dl	65	
13	13m	Н	Н	NHAc	CH ₃ CN	NaOAc	endo-11bm	а	
^a No convers	^a No conversion in methanol or acetonitrile under either basic or base-free conditions.								

Table 4. Scope and Limitations of Matsuda-Heck Couplings with 9a

entry	13	\mathbb{R}^2	\mathbb{R}^3	solvent	base	ratio ^a endo: exo	product	yield b (%)
1	13a	Н	OMe	CH ₃ CN	NaOAc	2.7:1.0	11aa	85
2	13a	Н	OMe	toluene	NaOAc	2.0:1.0	11aa	84
3	13a	Н	OMe	CH ₃ OH		1.0:6.6	11aa	73
4	13b	H	ОН	CH ₃ OH		1.0:6.0	11ab	76
5	13d	Br	OMe	CH ₃ CN	NaOAc	1.0:1.4	11ad	96
6	13e	Br	ОН	CH ₃ OH	NaOAc	10.0:1.0	11ae	44
7	13e	Br	ОН	CH ₃ OH		10.0:1.0	11ae	32
8	13f	NO_2	OMe	CH ₃ OH		2.0:1.0	11af	94
9	13g	NO_2	ОН	CH ₃ OH		2.0:1.0	11ag	99

"Determined by 1H NMR spectroscopy of the crude reaction mixture. "Combined isolated yield of endo- and exo-products."

investigated so far, all arylations of the five-membered *exo*-methylene lactone **9a** proceed with formation of both *endo*- and *exo*-isomers in varying ratios (Table 4). For each experiment, the *endo/exo* ratio was determined by ¹H NMR spectroscopy of the crude reaction mixture prior to chromatographic purification. *Endo*- and *exo*-isomers are conveniently distinguished by the chemical shift values for the olefinic proton and by the presence of a triplet integrating for two protons at ca. 3 ppm for the H³ protons of the *exo*-isomers. An NOE interaction between this methylene group and the *ortho*-protons of the aromatic substituent is indicative for the assigned *E*-configuration. Our configurational assignment differs from

that made by Arcadi et al. in their pioneering study, 10,11 who stated that the double bonds of their *exo*-products are exclusively *Z*-configured. However, a comparison of the NMR-spectroscopic data obtained by us for the *p*-methoxybenzylidene lactone *E-exo-11aa* and those reported by Arcadi et al. for *Z-exo-11aa* revealed that both data sets are identical, which suggests that the original assignment of a *Z*-configuration might be erroneous. As mentioned in the Introduction, Arcadi et al. proposed that acetate bases steer the β -hydride elimination toward the *endo-\beta*-H leading to a preferred formation of *endo*-butenolides. We found that the Matsuda–Heck coupling of **9a** and **13a** in acetonitrile in the presence of

acetate resulted in the formation of 11aa in a 2.7:1.0 ratio of endo- and exo-products (entry 1). In base-free methanol, the ratio is inverted to 1.0:6.6 (endo-11aa/exo-11aa), which seems to corroborate Arcadi's hypothesis that the base plays a crucial role for the regioselectivity (entry 3). Replacing acetonitrile by toluene under basic conditions (entry 2) gives 11aa in a comparable yield and very similar endo/exo ratio, which suggests that the base, rather than the solvent, is the dominant selectivity-controlling factor.

We found a comparable selectivity toward the *exo*-isomer for the phenoldiazonium salt **13b** in methanol under base free conditions (entry 4). However, as pointed out by Arcadi et al., the electronic effects of substituents at the arene moiety have a strong influence on the regioselectivity, with electron-with-drawing groups favoring *endo-\beta*-H-elimination. This might explain why even in the absence of acetate no pronounced selectivity toward the *exo*-isomer was observed for the diazonium salts **13d-g** (bearing an additional bromo or nitro substituent), although one should not overestimate the electronic effects of these substituents as they are located in a *meta*-position relative to the oxidative addition site.

Arcadi et al. showed that a homogeneous product can be obtained by hydrogenation of the crude endo/exo-mixtures with Pd/C.¹¹ While Arcadi's two-step sequence of Heck reaction and hydrogenation requires orthogonal solvents for each step (DMF and ethyl acetate, respectively), we sought to develop a one-pot, single-catalyst sequence by exploiting the advantages of arene diazonium salts as coupling partners. In particular, Matsuda-Heck reactions can be run in the absence of bases or ligands that might have detrimental effects on a subsequent hydrogenation step, they proceed normally at ambient temperature, which should be beneficial for catalyst longevity, and methanol is a suitable solvent for both coupling and hydrogenation step. After Matsuda-Heck couplings of 13a and 13b were run with α -methylene- γ -butyrolactone 9a in methanol under either basic or base-free conditions, activated charcoal was added once the evolution of nitrogen had ceased, and the reaction mixture was flushed with hydrogen and stirred under hydrogen at atmospheric pressure for 12 h. The expected α -benzylated γ -butyrolactones **16aa** and **16ab** were isolated in high yields (Scheme 6).

Scheme 6. One-Pot, Single-Catalyst Matsuda—Heck Coupling/Hydrogenation Sequence

Analysis of the Regioselectivity of the β -H-Elimination Step Based on DFT Calculations. The strikingly different selectivities observed for Matsuda—Heck arylations of five- and six-membered *exo*-methylene lactones might be of thermodynamic or kinetic origin. Thermodynamic control would require a feasible "post-Mizoroki—Heck" double-bond isomerization mechanism, ⁵⁸ e.g., via a sufficiently stable Pd hydride species [H–Pd–X] (4, Scheme 1) originating from the β -H-

elimination step. This Pd hydride might be capable of isomerizing the kinetic product into the thermodynamic one by hydropalladation $-\beta$ -H-elimination steps. Destabilization of Pd hydrides, e.g., by trapping stabilizing iodide ligands with silver⁵⁹ or thallium salts,⁶⁰ is hence a measure to suppress unwanted post-Mizoroki-Heck double-bond migration reactions. With arene diazonium tetrafluoroborates as arylating agents, strongly coordinating halide ligands are absent from the outset and the catalytically relevant intermediates are supposedly cationic, 47,61 which means that rather unstable Pd hydride species have to be expected. It is, for instance, in line with this assumption that no subsequent isomerization of the double bond has been observed in Matsuda-Heck reactions with cyclic enol ethers, while the analogous reactions with aryl iodides were found to be prone to post-Mizoroki-Heck reactions. 62 In light of these considerations, it appears unlikely that the different regioselectivities observed for five- and sixmembered exo-methylene lactones result from a Pd hydride catalyzed double-bond migration following the Matsuda-Heck reaction. To corroborate this supposition, the relative stabilities of all possible Matsuda-Heck products resulting from the reaction of phenol diazonium salt 13b with 9a and 9b, respectively, were determined by calculating their Gibbs free energy values using DFT methods (Figure 1).

Both *exo-Z*-isomers were found to be clearly the least stable products. In Figure 1 their Gibbs free energy is set to 0, and the free energies of the isomeric *endo-* and *exo-E-*products are denoted relative to *exo-Z-*11ab and *exo-Z-*11bb, respectively. For both ring sizes, the order of stability is identical: $exo-E > endo \gg exo-Z$. This underlines that the strikingly different

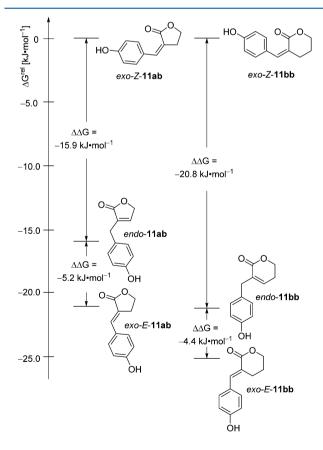


Figure 1. Calculated relative Gibbs free energies of isomers of 11ab and 11bb.

product distributions observed for Matsuda-Heck reactions of five- and six-membered α -methylene lactones most likely do not originate from thermodynamic but from kinetic control. It should be noted in this context that a single report exists which describes selective Rh-catalyzed exo-to-endo double-bond migrations, including conversions of exo-E-11aa and exo-E-11ba to endo-11aa and endo-11ba, respectively, which apparently contradicts our assumption.⁸⁴ Although alternative β -H-elimination scenarios such as *trans-\beta*-H-elimination have been proposed in cases of conformationally constrained Pd σ complexes, we assumed that a β -H-elimination will preferably proceed via the syn-mechanism, provided that a syn- β -H is available. To gain a deeper understanding of the different β -Helimination pathways, the cationic Pd σ -complexes 10 (Scheme 1) resulting from the migratory insertion into the exo-double bonds of 9a and 9b, respectively, were analyzed using DFT methods. For the Pd an oxidation state of +II and the coordination of two molecules of methanol was assumed. The $Pd-C^{\alpha}-C^{\beta}-Ar$ dihedral angles were then locked to 0°, and geometry optimizations were performed. The structures shown in Figure 2 mirror the situation immediately after the migratory insertion step (Figure 2).

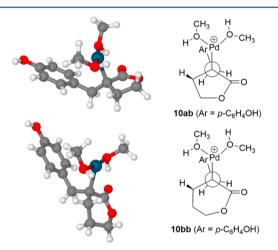


Figure 2. Geometry-optimized structures of Pd σ -complexes **10ab** and **10bb** with a dihedral angle $[Pd-C^{\alpha}-C^{\beta exo}-Ar]=0^{\circ}$.

Starting from these eclipsed-conformations, which are highest in energy, the aryl moiety was rotated stepwise around the $C^{\alpha}-C^{\beta exo}$ bond between 0° and 360° . Geometry optimizations were performed for 36 conformers, and the differences in energy relative to the starting conformation and the dihedral angles $[Pd-C^{\alpha}-C^{\beta endo}-H]$ were calculated for these 36 conformers for each Pd σ -complex 10ab and 10bb. The results for the five-membered lactone structure 10ab are shown in Figure 3. The first relevant conformation arises after rotation of 120° around the $C^{\alpha}-C^{\beta exo}$ bond. A β -H^{exo} elimination from this conformer would lead to exo-Z-11ab, but this conformer is as high in energy as the starting conformer. exo-Z-11ab is obviously not only thermodynamically (see Figure 1) but also kinetically strongly disfavored. Rotating the aryl substituent by a further 120° (dihedral angle $[Pd-C^{\alpha}-C^{\beta exo}-Ar] = 240^{\circ})$ leads to another syn- β -H^{exo} arrangement, which is energetically much more favorable than the structure at 120° . $syn-\beta$ -H-elimination from this conformer would lead to exo-E-11ab, which is indeed the main product observed in this Matsuda-Heck reaction (see Table 4). As a

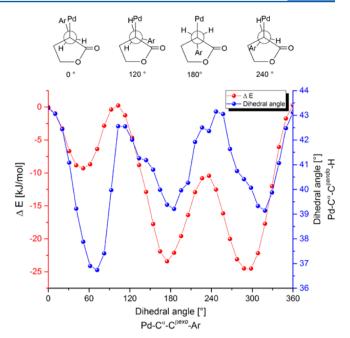


Figure 3. $[Pd-C^{\alpha}-C^{\beta endo}-H]$ dihedral angle (blue curve) and relative energy (red curve) as a function of the $[Pd-C^{\alpha}-C^{\beta exo}-Ar]$ dihedral angle for Pd σ -complex **10ab**.

measure for the probability of a $syn-\beta$ -H^{endo} elimination, we considered the endo-dihedral angle $[Pd-C^{\alpha}-C^{\beta endo}-H]$, which varies from 37° to 44°, depending on the exo-dihedral angle $[Pd-C^{\alpha}-C^{\beta exo}-Ar]$ (blue curve in Figure 3). We reasoned that for a $syn-\beta$ -H^{endo} elimination to proceed efficiently two conditions have to be met: the endo-dihedral angle needs to be sufficiently small and the respective conformer should be energetically favorable. The smallest endo-dihedral angle (36.5°) coincides with the gauche conformation at 60° (exo-dihedral angle), but this conformer is, although at a local minimum, still rather high in energy. On the other hand, the two energetically most favorable conformers at exo-dihedral angles of 180° and 300° have larger endo-dihedral angles of ca. 39° (Figure 3).

In the case of the six-membered lactone structure 10bb, the analogous analysis revealed some striking differences (Figure 4). Both conformers of Pd σ -complex 10bb with a syn- β -H^{exo} arrangement (at 120° and at 240°, which would hypothetically give exo-Z- and exo-E-11bb, respectively, upon β -H-elimination) are equally high in energy and only marginally more stable than the starting conformer. In contrast, the conformer at 240° of the five-membered lactone structure 10ab, which is the precursor for the exo-E-product, is ca. 10 kJ·mol⁻¹ lower in energy than the respective starting conformer. This might explain why no exo-E-products were observed for Matsuda-Heck reactions with six-membered α -methylene lactones. The pronounced preference for the formation of endo-products can be explained by considering the endo-dihedral angle (blue curve in Figure 4). Compared to the five-membered lactone structures, this crucial dihedral angle is generally smaller, and an energetically favorable conformer exists at 180°, which nearly coincides with the smallest calculated endo-dihedral angle of ca. 22°. For the two other local energy minima endo-dihedral angles larger than 40° were calculated. It appears unlikely that a syn- β - H^{endo} -elimination will occur from these conformers.

Total Synthesis of Anemarcumarin A. Several natural products with a 3-benzylated coumarin pattern have been

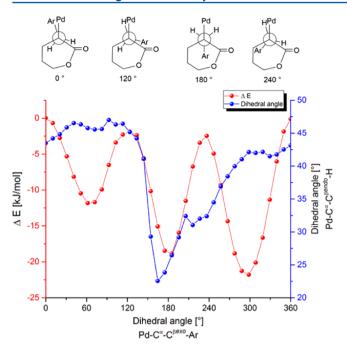


Figure 4. [Pd- C^{α} - $C^{\beta endo}$ -H] dihedral angle (blue curve) and relative energy (red curve) as a function of the [Pd- C^{α} - $C^{\beta exo}$ -Ar] dihedral angle for Pd σ -complex **10bb**.

isolated from plants. 63,64 Examples are sarcandracoumarin, a weakly active cytotoxic compound isolated from the plant *Sarcandra glabra*, 65 or anemarcoumarin A, which has been isolated from the rhizomes of *Anemarrhena asphodeloides*, a medicinal plant native to Korea, China, and Japan. 66 Synthetic 3-benzylated coumarins have recently been evaluated for various medicinal chemistry purposes, e.g., as selective CB₂ agonists, $^{67-69}$ as inhibitors of TNF- α , 70 or as inhibitors of estrogen receptors (Figure 5). A previously described synthetic route to these 3-benzylcoumarins proceeds through umpolung—domino reactions of salicylic aldehydes and enals. 72

Anemarcoumarin A

Selective cannabinoid receptor (CB₂) agonist PSB-SB1204

OMe

HO

OH

HO

OH

HO

OH

HO

OH

OH

HO

OH

NMe₂

OH

Inhibitor of tumor necrosis factor (TNF-
$$\alpha$$
); anti-inflammatory

Figure 5. Examples for 3-benzylated coumarins as potential target structures.

We applied the Matsuda—Heck conditions developed herein the synthesis of anemarcoumarin A from umbelliferone (17). Umbelliferone was first hydrogenated and then protected as TBS ether 19. Application of the conditions used for *exo*methylenation of lactones 12 as described in Scheme 2 failed in this particular case and resulted in a complex mixture of products. However, deprotonation of 19 with in situ generated LDA, followed by addition of Eschenmoser's salt⁷³ and methyl iodide, furnished the required *exo*-methylene coumarine 20.⁷⁴

Matsuda—Heck arylation of **20** with phenol diazonium salt **13b** led to an inseparable mixture of products, which contained only minor amounts of the desired coupling product. Interestingly, the Matsuda—Heck coupling worked satisfactorily with **21**, obtained after desilylation of **20**, under otherwise identical conditions and furnished anemarcoumarin A in six steps from umbelliferone in an overall yield of 28% (Scheme 7).

Scheme 7. Total Synthesis of Anemarcoumarin A from Umbelliferone

CONCLUSIONS

In summary, we report conditions for the base- and ligand-free Heck-type arylation of α -methylene lactones with arene diazonium salts. While mostly poor levels of regioselectivity were observed for α -methylene- γ -butyrolactone, the homologous six-membered α -methylene lactone and the analogous lactam react to the Matsuda-Heck products with perfect endoselectivity. Based on DFT calculations, we suggest that the high levels of endo-selectivity observed for six-membered substrates result from kinetic rather than thermodynamic control. We found that for the Pd σ -complex an energetically favorable conformation coincides with a comparatively small $[Pd-C^{\alpha} C^{\beta endo}$ -H] dihedral angle, which is not the case for the fivemembered lactone. On the other hand, a conformer with a Pd- $H^{\beta exo}$ syn arrangement which is lower in energy and leads to an exo-E-product was observed for the five-membered lactone. An endo-selective Matsuda-Heck arylation of an α -methylene coumarin was finally applied to the synthesis of the plant natural product anemarcoumarin A.

■ EXPERIMENTAL SECTION

General Methods. All experiments were conducted in dry reaction vessels under an atmosphere of dry nitrogen. Solvents were purified by standard procedures. 1 H NMR spectra were obtained at 300 MHz in CDCl₃ with CHCl₃ (δ = 7.26 ppm) as an internal standard. Coupling constants are given in hertz. 13 C NMR spectra were recorded at 75 MHz in CDCl₃ with CDCl₃ (δ = 77.1 ppm) as an internal standard. Whenever the solubility of the sample or signal separation were insufficient in CDCl₃, it was replaced by one of the following solvents: DMSO- d_6 (DMSO- d_5 as internal standard for 14 H NMR spectroscopy, δ = 2.50 ppm, DMSO- d_6 as internal standard for 13 C NMR spectroscopy, δ = 39.5 ppm); C_6D_6 (C_6D_5 as internal standard for

 1 H NMR spectroscopy, δ = 7.16 ppm, C_6D_6 as internal standard for 13 C NMR spectroscopy, δ = 128.1 ppm); acetone- d_6 (acetone- d_5 as internal standard for 1 H NMR spectroscopy, δ = 2.05 ppm, CD_3COCD_3 as internal standard for 13 C NMR spectroscopy, δ = 29.8 ppm); methanol- d_4 (CD_2HOD as internal standard for 14 H NMR spectroscopy, δ = 3.31 ppm, CD_3OD as internal standard for 13 C NMR spectroscopy, δ = 49.0 ppm). IR spectra were recorded as ATR-FTIR spectra. Wavenumbers (ν) are given in cm $^{-1}$. The peak intensities are defined as strong (s), medium (m) or weak (w). Lowand high-resolution mass spectra were obtained by EI-TOF or ESI-TOF. The following arene diazonium salts were synthesized from anilines by diazotation or from acetanilides through the deacetylation—diazotation sequence, according to previously published procedures: 13a, 29 13b, 14 13c, 29 13d, 29 13

Computational Methods. All DFT calculations were performed with the B3LYP density functional^{75–77} as implemented in the ORCA⁷⁸ program package. We used the def2-TZVP basis set⁷⁹ for all atoms and a Stuttgart–Dresden effective core potential (SD(28, MWB))⁸⁰ for palladium. To further account for the methanol solvent effects, the conductor-like screening model (COSMO)⁸¹ was used. A frequency calculation was performed for the stationary points to calculate the thermochemical properties.

3-Methylenetetrahydro-2H-pyran-2one (9b).³⁶ A suspension of NaH (60 wt % dispersion in mineral oil, 1.12 g, 28.2 mmol) in THF (100 mL) was cooled to 0 °C, and a solution of 12b (1.70 g, 17.1 mmol) and diethyl oxalate (5.10 g, 35.1 mmol) in THF (100 mL) was added dropwise. After addition was complete, ethanol (4.5 mL) was added, and the reaction mixture was warmed to ambient temperature and stirred for 4 h. After the mixture was cooled to 0 °C, a solution of K₂CO₃ (9.60 g, 70.0 mmol) in water (14.1 mL) and formaldehyde (37 wt % aq solution, 18.9 g) were added, and the reaction mixture was stirred for 0.25 h at this temperature. After this time, the mixture was diluted with brine (50 mL). The layers were separated, and the aqueous layer was extracted twice with diethyl ether (50 mL each). The combined organic extracts were washed with brine, dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures of increasing polarity as eluent, to furnish 9b (1.50 g, 13.4 mmol, 78%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 6.41–6.37 (m, 1H), 5.55–5.51 (m, 1H), 4.35 (t, J = 5.4 Hz, 2H), 2.68–2.58 (m, 2H), 1.99–1.84 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 165.6, 134.2, 128.3, 69.8, 28.2, 23.3; IR (ATR) ν 1714 (s), 1624 (w), 1399 (m), 1294 (s), 1143 (s); HRMS (EI) calcd for C₆H₈O₂ [M⁺] 112.0524, found 112.0528.

6-Methyl-3-methylenetetrahydro-2H-pyran-2one (9c).³⁷ Following the procedure for 9b, 12c (2.00 g, 17.1 mmol) was converted to 9c (1.50 g, 12.0 mmol, 70%): colorless liquid; ¹H NMR (300 MHz, C_6D_6) δ 6.41–6.37 (m, 1H), 5.10–5.07 (m, 1H), 3.91–3.78 (m, 1H), 2.10 (dm, J = 16.3 Hz, 1H), 1.96 (ddm, J = 16.4, 12.2 Hz, 1H), 1.20 (dm, J = 13.9 Hz, 1H), 1.10–0.95 (m, 1H), 0.96 (d, J = 6.4 Hz, 3H); ¹³C NMR (75 MHz, C_6D_6) δ 164.4, 134.5, 126.4, 76.0, 29.8, 27.0, 21.2; IR (ATR) ν 1714 (s), 1387 (m), 1295 (s), 1170 (m), 1129 (s).

Ethyl 1-Benzylpiperdine-3-carboxylate (N-Bn-12d). A solution of ethyl nipecotate (12d, 7.90 g, 50.0 mmol), NEt₃ (14.10 mL, 100.0 mmol), and benzyl bromide (6.60 mL, 55.0 mmol) in acetonitrile (30 mL) was heated to 80 °C until full conversion of the starting material. The solvent was evaporated, and aq NaOH (1 M) was added (pH = 12). The residue was extracted three times with CH₂Cl₂ (50 mL each). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures as eluent, to furnish N-Bn12d (12.30 g, 50.0 mmol, quant): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.43–7.12 (m, 5H), 4.13 (q, J = 7.1 Hz, 2H), 3.58 (d, J = 13.2 Hz, 1H), 3.50 (d, J = 13.2 Hz, 1H), 2.97 (dm, J = 11.0 Hz, 1H), 2.74 (dm, J = 11.3 Hz, 1H), 2.66-2.54 (m, 1H), 2.26 (dd, J = 10.5,10.5 Hz, 1H), 2.07 (ddd, J = 10.8, 10.8, 2.8 Hz, 1H), 2.00–1.88 (m, 1H), 1.82-1.70 (m, 1H), 1.70-1.40 (m, 2H), 1.25 (t, J = 7.1 Hz, 3H); 13 C NMR (75 MHz, CDCl₃) δ 174.3, 138.4, 129.0, 128.2, 127.0, 63.3, 60.2, 55.4, 53.6, 41.9, 27.0, 24.6, 14.2; IR (ATR) ν 2941 (w), 1729 (s),

1189 (m), 1154 (m); HRMS (EI) calcd for $C_{15}H_{21}NO_2$ [M⁺] 247.1572, found 247.1576.

1-Benzyl-3-methylenepiperidin-2-one (9d).82 A solution of N-Bn-12d (9.50 g, 38.4 mmol) and NaOH (1.70 g, 42.5 mmol) in water (18 mL) and methanol (350 mL) was stirred at ambient temperature for 12 h. After this time, the solvent was evaporated, and the residue was suspended in toluene and evaporated again. To the dry residue were added acetanhydride (370 mL) and NEt₃ (54 mL), and the reaction mixture was heated to 90 °C for 12 h. All volatiles were evaporated, and water (100 mL) and CH₂Cl₂ (100 mL) were added. The aqueous layer was extracted three times with CH₂Cl₂ (100 mL). The combined organic extracts were washed with brine (50 mL), dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures of increasing polarity as eluent, to furnish 9d (5.40 g, 26.8 mmol, 70%): yellowish liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.37-7.21 (m, 5H), 6.28 (ddd, I = 1.8, 1.8, 1.8 Hz, 1H), 5.31 (dd, I = 1.8, 1.8, 1.8 Hz, 1H), 4.66 (s, 2H), 3.32-3.24 (m, 2H), 2.67-2.49 (m, 2H), 1.99-1.74 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 164.4, 137.9, 137.4, 128.7, 128.2, 127.4, 122.0, 50.8, 47.9, 30.3, 23.2; IR (ATR) ν 2932 (w), 1656 (s), 1610 (s), 1486 (m), 1451 (m), 1222 (m); HRMS (EI) calcd for C₁₃H₁₅NO [M⁺] 201.1154, found 201.1158.

4-Hydroxy-2-(methoxycarbonyl)benzenediazonium Tetrafluoroborate (13l). A suspension of 14l (1.00 g, 6.0 mmol), tetrafluoroborate (50 wt % in water, 1.20 mL, 9.6 mmol), water (0.74 mL) and 2-propanol (1.00 mL) was stirred for 0.5 h at ambient temperature. The resulting solution was then cooled to 0 °C, and NaNO₂ (0.82 g, 12.0 mmol) was added in small portions. The suspension was stirred for 0.5 h at 0 °C and filtered through a Büchner funnel. The solid was washed with cold water (15 mL), ethanol, and then diethyl ether and dried in vacuum to give 13l (0.71 g, 2.7 mmol, 42%): grayish solid; ¹H NMR (300 MHz, DMSO- d_6) δ 8.32 (d, J = 9.4 Hz, 1H), 7.16 (d, J = 2.3 Hz, 1H), 6.90 (dd, J = 9.4, 2.3 Hz, 1H), 3.93 (s, 3H); ¹³C NMR (75 MHz, DMSO- d_6) δ 174.6, 162.4, 136.7, 132.9, 123.9, 123.6, 87.4, 53.6; IR (KBr-disc) ν 3320 (w), 2181 (s), 1735 (s), 1709 (s), 1437 (m), 1292 (s); HRMS (ESI) calcd for $C_8H_7N_2O_3$ [M⁺] 179.0457, found 179.0454

General Procedure 1: Matsuda—Heck Couplings of 9a,b under Base-Free Conditions. The appropriate arene diazonium salt 13 (0.40 mmol) and $Pd(OAc)_2$ (4.5 mg, 5.0 mol %) were suspended in the corresponding solvent as indicated in Table 2 (4.0 mL). α-Methylene-γ-butyrolactone 9a (78 mg, 0.80 mmol) or α-methylene-δ-valerolactone 9b (90 mg, 0.80 mmol) was added, and the reaction mixture was stirred at ambient temperature for 12 h. The solvent was evaporated, and the residue was purified by column chromatography on silicia using hexane–MTBE mixtures of increasing polarity as eluent.

General Procedure 2: Matsuda—Heck Couplings of 9a,b under Basic Conditions. The appropriate arene diazonium salt 13 (0.40 mmol), $Pd(OAc)_2$ (4.5 mg, 5.0 mol %), and NaOAc (98 mg, 1.20 mmol) were suspended in the corresponding solvent as indicated in Table 2 (4.0 mL). α-Methylene-γ-butyrolactone 9a (78 mg, 0.80 mmol) or α-methylene-δ-valerolactone 9b (90 mg, 0.80 mmol) was added, and the reaction mixture was stirred at ambient temperature for 12 h. All volatiles were evaporated, and the residue was partitioned between water (10 mL) and MTBE (10 mL). The aqueous layer was separated and extracted with MTBE (3 times, 10 mL for each extraction). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica using hexane—MTBE mixtures of increasing polarity as eluent.

3-(4-Methoxybenzyl)furan-2(5H)-one (endo-11aa)¹¹ and (E)-3-(4-Methoxybenzylidene)dihydrofuran-2(3H)-one (exo-11aa).⁸³ Following general procedure 2 (solvent: acetonitrile), 9a (78 mg, 0.80 mmol) and 13a (89 mg, 0.40 mmol) were converted to a 2.7:1.0 mixture (determined by ¹H NMR analysis of the crude mixture) of endo- and exo-11aa (70 mg, 0.34 mmol, 85%). Following general procedure 1 (solvent: methanol), 9a (78 mg, 0.80 mmol) and 13a (89 mg, 0.40 mmol) were converted to a 1.0:6.6 mixture (determined by ¹H NMR analysis of the crude mixture) of endo- and exo-11aa (60 mg,

0.29 mmol, 73%). NMR data of *endo-***11a**a: ¹H NMR (300 MHz, CDCl₃) δ 7.13 (d, J = 8.5 Hz, 2H), 6.92 (pent, J = 1.8 Hz, 1H), 6.83 (d, J = 8.5 Hz, 2H), 4.72 (q, J = 1.9 Hz, 2H), 3.76 (s, 3H), 3.50 (q, J = 1.9 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 174.0, 158.5, 145.5, 134.5, 129.9, 129.4, 114.1, 70.3, 55.3, 31.0. NMR data of *exo-***11aa**: ¹H NMR (300 MHz, CDCl₃) δ 7.47 (t, J = 2.9 Hz, 1H), 7.42 (d, J = 8.7 Hz, 2H), 6.93 (d, J = 8.7 Hz, 2H), 4.41 (t, J = 7.3 Hz, 2H), 3.82 (s, 3H), 3.17 (td, J = 7.3, 2.9 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 160.9, 136.3, 131.8, 127.4, 120.7, 114.4, 65.4, 55.4, 27.4; HRMS (ESI) calcd for $C_{12}H_{13}O_3$ [M + H]⁺ 205.0865, found 205.0865.

3-(4-Hydroxybenzyl)furan-2(5H)-one (endo-11ab) and (E)-3-(4-Hydroxybenzyl)furan-2(5H)-one (exo-11ab). ⁸³ Following general procedure 1 (solvent: methanol), 9a (78 mg, 0.80 mmol) and 13b (83 mg, 0.40 mmol) were converted to a 1.0:6.0 mixture (determined by ¹H NMR analysis of the crude mixture) of *endo-* and *exo-11ab* (58 mg, 0.30 mmol, 76%). NMR data of *endo-11ab*: ¹H NMR (300 MHz, DMSO- d_6) δ 9.32 (s (br.), 1H), 7.28 (pent, J = 1.6 Hz, 1H), 7.03 (d, J = 8.4 Hz, 2H), 6.69 (d, J = 8.4 Hz, 2H), 4.80 (q, J = 1.8 Hz, 2H), 3.40 (s, 2H); ¹³C NMR (75 MHz, DMSO- d_6) δ 175.0, 156.0, 148.0, 132.3, 129.8, 128.1, 115.4, 70.7, 30.2. NMR data of *exo-11ab*: ¹H NMR (300 MHz, DMSO- d_6) δ 10.14 (s (br.), 1H), 7.46 (d, J = 8.6 Hz, 2H), 7.31 (t, J = 2.6 Hz, 1H), 6.87 (d, J = 8.6 Hz, 2H), 4.37 (t, J = 7.3 Hz, 2H), 3.15 (td, J = 7.2, 2.6 Hz, 2H); ¹³C NMR (75 MHz, DMSO- d_6) δ 172.6, 159.3, 135.2, 132.3, 125.8, 120.9, 116.1, 65.5, 27.0; HRMS (ESI) calcd for $C_{11}H_{11}O_3$ [M + H]⁺ 191.0708, found 191.0709.

3-(3-Bromo-4-methoxybenzyl)furan-2(5H)-one (endo-**11ad**) and (E)-3-(3-Bromo-4-methoxy-benzylidene)dihydrofuran-2(3H)-one 3-(3-Bromo-4-methoxybenzyl)furan-2(5H)-one (exo-11ad). Following general procedure 2 (solvent: acetonitrile), 9a (78 mg, 0.80 mmol) and 13d (120 mg, 0.40 mmol) were converted to a 1.0:1.4 mixture (determined by ¹H NMR analysis of the crude mixture) of endo- and exo-11ad (108 mg, 0.38 mmol, 96%). NMR data of endo-11ad: ¹H NMR (300 MHz, CDCl₃) δ 7.43–7.34 (m, 1H, signal overlap with endo isomer), 7.14 (dd, J = 8.3, 1.8 Hz, 1H), 7.00 (pent, J = 1.7 Hz, 1H), 6.83 (d, J = 8.4 Hz, 1H), 4.75 (q, J = 1.8 Hz, 2H), 3.84 (s, 3H), 3.51–3.47 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 173.8, 154.7, 145.9, 133.7, 133.5, 130.9, 129.0, 112.1, 111.6, 70.3, 56.3, 30.5. NMR data of exo-11ad: ¹H NMR (300 MHz, CDCl₃) δ 7.64 (d, J = 1.8 Hz, 1H), 7.43-7.34 (m, 2H, signal overlap with endo isomer), 6.94 (d, J =8.3 Hz, 1H), 4.43 (t, J = 7.2 Hz, 2H), 3.91 (s, 3H), 3.18 (td, J = 7.3, 2.9 Hz, 2H); 13 C NMR (75 MHz, CDCl₃) δ 172.4, 156.9, 134.6, 134.3, 131.1, 128.6, 122.3, 112.2, 112.0, 65.4, 56.4, 27.2; HRMS (ESI) calcd for C₁₂H₁₂⁷⁹BrO₃ [M + H]⁺ 282.9970, found 282.9957.

3-(3-Bromo-4-hydroxybenzyl)furan-2(5H)-one (endo-11ae) and (E)-3-(3-Bromo-4-hydroxybenzylidene)dihydrofuran-2(3H)-one (exo-11ae). Following general procedure 2 (solvent: methanol), 9a (78 mg, 0.80 mmol) and 13e (115 mg, 0.40 mmol) were converted to a 10.0:1.0 mixture (determined by ¹H NMR analysis of the crude mixture) of endo- and exo-11ae (47 mg, 0.18 mmol, 44%). Following general procedure 1 (solvent: methanol), 9a (78 mg, 0.80 mmol) and 13e (115 mg, 0.40 mmol) were converted to a 10.0:1.0 mixture (determined by ¹H NMR analysis of the crude mixture) of endo- and exo-11ae (34 mg, 0.13 mmol, 32%). Selected ¹H NMR data of exo-**11ae**: 1 H NMR (300 MHz, DMSO- d_{6}) δ 10.14 (s (br.), 1H), 7.77 (d, J= 2.0 Hz, 1H), 7.48 (dd, J = 8.6, 2.0 Hz, 1H), 7.31 (t, J = 2.7 Hz, 1H),4.40 (t, J = 7.2 Hz, 2H), 3.20 (td, J = 7.2, 2.7 Hz, 2H). NMR data of endo-11ae: 1 H NMR (300 MHz, DMSO- d_{6}) δ 10.09 (s, 1H), 7.38– 7.34 (m, 2H), 7.05 (dd, J = 8.3, 2.0 Hz, 1H), 6.88 (d, J = 8.3 Hz, 1H), 4.83 (q, J = 1.7 Hz, 2H), 3.43 (s, 2H); ¹³C NMR (75 MHz, DMSO d_6) δ 173.7, 152.6, 148.3, 132.8, 131.6, 130.0, 129.0, 116.3, 109.1, 70.6, 29.6; HRMS (EI) calcd for $C_{11}H_9^{79}BrO_3$ [M⁺] 267.9735, found 267.9728.

3-(4-Methoxy-3-nitrobenzyl)furan-2(5H)-one (endo-11af) and (E)-3-(4-Methoxy-3-nitrobenzylidene)dihydrofuran-2(3H)-one (exo-11af). Following general procedure 1 (solvent: methanol), 9a (78 mg, 0.80 mmol) and 13f (108 mg, 0.40 mmol) were converted to a 2.0:1.0 mixture (determined by 1 H NMR analysis of the crude mixture) of endo- and exo-11af (93 mg, 0.38 mmol, 94%). NMR data of endo-11af: 1 H NMR (300 MHz, CDCl₃) δ 7.74 (d, J = 1.9 Hz, 1H), 7.49 (dd, J = 8.9, 2.0 Hz, 1H), 7.09—7.05 (m, 2H), 4.82 (q, J = 1.7 Hz, 2H), 3.97 (s,

3H), 3.63 (s, 2H); 13 C NMR (75 MHz, CDCl₃) δ 173.5, 152.0, 146.0, 134.8, 125.8, 113.9, 70.3, 56.6, 30.6, other signals cannot be unambigously assigned due to signal overlap. NMR data of *exo-***11af**: 1 H NMR (300 MHz, CDCl₃) δ 8.03 (d, J = 1.8 Hz, 1H), 7.69 (dd, J = 8.8, 1.9 Hz, 1H), 7.51–7.48 (m, 1H), 7.19 (d, J = 8.8 Hz, 1H), 4.52 (t, J = 7.2 Hz, 2H), 4.04 (s, 3H), 3.28 (td, J = 7.2, 2.8 Hz, 2H); 13 C NMR (75 MHz, CDCl₃) δ 65.4, 56.8, 27.2, other signals cannot be unambigously assigned due to signal overlap; HRMS (ESI) calcd for $C_{12}H_{12}NO_5$ [M + H] $^+$ 250.0715, found 250.0702.

3-(4-Hvdroxy-3-nitrobenzyl)furan-2(5H)-one (endo-**11aa**) and (E)-3-(4-Hydroxy-3-nitrobenzylidene)dihydrofuran-2(3H)-one (exo-11ag). Following general procedure 1 (solvent: methanol), 9a (78 mg, 0.80 mmol) and 13g (101 mg, 0.40 mmol) were converted to a 2.0:1.0 mixture (determined by 1H NMR analysis of the crude mixture) of endo- and exo-11ag (93 mg, 0.40 mmol, 99%). NMR data of endo-11ag: ¹H NMR (300 MHz, CDCl₃) δ 10.48 (s, 1H), 7.97 (d, J = 1.8 Hz, 1H), 7.52 (dd, J = 8.6, 1.9 Hz, 1H), 7.13 (d, J = 8.6 Hz, 1H),7.13–7.10 (m, 1H), 4.82 (q, J = 1.7 Hz, 2H), 3.62 (s, 2H); ¹³C NMR (75 MHz, CDCl₂) δ 173.5, 154.0, 146.2, 138.4, 124.7, 120.4, 70.4, 30.6, other signals cannot be unambigously assigned. NMR data of exo-11ag: ¹H NMR (300 MHz, CDCl₃) δ 10.71 (s, 1H), 8.26 (d, J = 1.6Hz, 1H), 7.74 (dd, J = 8.7, 1.9 Hz, 1H), 7.48 (t, J = 2.8 Hz, 1H), 7.25(d, J = 8.8 Hz, 1H), 4.52 (t, J = 7.2 Hz, 2H), 3.28 (td, J = 7.2, 2.8 Hz,2H); ^{13}C NMR (75 MHz, CDCl $_3$) δ 171.9, 155.6, 133.6, 127.4, 124.6, 120.9, 65.4, 27.2, other signals can not be unambigously assigned; HRMS (ESI) calcd for $C_{11}H_{10}NO_5$ [M + H]⁺ 236.0559, found 236.0560.

3-(4-Methoxybenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11ba). ⁸⁴ Following general procedure 2 (solvent: acetonitrile), 9b (112 mg, 1.00 mmol) and 13a (111 mg, 0.50 mmol) were converted to endo-11ba (106 mg, 0.49 mmol, 98%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.13 (d, J = 7.5 Hz, 2H), 6.85 (d, J = 7.4 Hz, 2H), 6.55–6.33 (m, 1H), 4.33 (t, J = 5.9 Hz, 2H), 3.81 (s, 3H), 3.56 (s, 2H), 2.44–2.34 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 165.3, 158.6, 140.2, 133.6, 130.7, 130.7, 114.4, 66.8, 55.7, 36.3, 24.8; IR (ATR) ν 1710 (s), 1510 (s), 1398 (m), 1243 (s); HRMS (ESI) calcd for $C_{13}H_{14}O_3$ [M + H]⁺ 219.1016, found 219.1016.

3-(4-Hydroxybenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bb). Following general procedure 1 (solvent: methanol), 9b (112 mg, 1.00 mmol) and 13b (105 mg, 0.50 mmol) were converted to endo-11bb (76 mg, 0.37 mmol, 74%): colorless solid; mp 106–108 °C; 1 H NMR (300 MHz, CDCl₃) δ 6.93 (d, J=8.5 Hz, 2H), 6.68 (d, J=8.5 Hz, 2H), 6.36 (t, J=4.3 Hz, 1H), 6.28 (s, 1H), 4.23 (t, J=6.3 Hz, 2H), 3.43 (d, J=1.1 Hz, 1H), 2.35–2.24 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 165.7, 154.8, 140.5, 133.2, 130.4, 129.9, 115.6, 66.6, 36.0, 24.5; IR (ATR) ν 3341 (bw), 1668 (s), 1513 (s), 1273 (m), 1117 (s); HRMS (EI) calcd for C₁₂H₁₂O₃ [M⁺] 204.0786, found 204.0776.

3-(4-(Benzyloxy)benzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bc). Following general procedure 2 (solvent: acetonitrile), 9b (90 mg, 0.80 mmol) and 13a (119 mg, 0.40 mmol) were converted to endo-11bc (78 mg, 0.26 mmol, 66%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 7.51–7.31 (m, 5H), 7.15 (d, J = 8.3 Hz, 2H), 6.95 (d, J = 8.6 Hz, 2H), 6.44 (t, J = 4.2 Hz, 1H), 5.02 (s, 2H), 4.36 (t, J = 6.2 Hz, 2H), 3.59 (s, 2H), 2.47–2.38 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 165.3, 157.9, 140.2, 137.5, 133.6, 131.0, 130.7, 129.0, 128.3, 127.9, 115.3, 70.5, 66.8, 36.3, 24.8; IR (ATR) ν 1711 (s), 1609 (m), 1509 (s), 1237 (s); HRMS (EI) calcd for $C_{19}H_{18}O_3$ [M $^+$] 294.1256, found 294.1245.

3-(3-Bromo-4-methoxybenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bd). Following general procedure 1 (solvent: methanol), 9b (90 mg, 0.80 mmol) and 13d (120 mg, 0.40 mmol) were converted to endo-11bd (98 mg, 0.33 mmol, 82%): colorless liquid; 1 H NMR (300 MHz, C_6D_6) δ 7.36 (d, J = 2.1 Hz, 1H), 7.01 (dd, J = 8.4, 2.1 Hz, 1H), 6.37 (d, J = 8.4 Hz, 1H), 5.69 (t, J = 4.2 Hz, 1H), 3.55 (t, J = 6.2 Hz, 2H), 3.39 (s, 2H), 3.25 (s, 3H), 1.50–1.29 (m, 2H); 13 C NMR (75 MHz, C_6D_6) δ 164.0, 155.1, 139.5, 134.1, 132.9, 132.9, 129.7, 112.3, 112.2, 65.8, 55.7, 36.2, 24.3; IR (ATR) ν 1712 (s), 1495 (s), 1400 (m), 1276 (s), 1255 (s), 1114 (s); HRMS (EI) calcd for $C_{13}H_{13}^{79}$ BrO₃ [M $^+$] 296.0048, found 296.0047.

3-(3-Bromo-4-hydroxybenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11be). Following general procedure 2 (solvent: methanol), 9b (112 mg, 1.00 mmol) and 13e (143 mg, 0.50 mmol) were converted to endo-11be (95 mg, 0.34 mmol, 68%): colorless liquid; ¹H NMR (300 MHz, acetone- d_6) δ 8.75 (s, 1H), 7.37 (d, J = 2.0 Hz, 1H), 7.07 (dd, J = 8.3, 2.1 Hz, 1H), 6.92 (d, J = 8.3 Hz, 1H), 6.70 (t, J = 4.3 Hz, 1H), 4.32 (t, J = 6.2 Hz, 2H), 3.48 (d, J = 1.1 Hz, 2H), 2.57–2.36 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 165.0, 153.3, 141.9, 134.1, 133.0, 132.9, 130.2, 117.1, 110.1, 67.1, 36.3, 25.1; IR (ATR) ν 3258 (m), 1648 (s), 1494 (m), 1416 (m), 1274 (s), 1115 (s); HRMS (ESI) calcd for $C_{12}H_{12}^{-79}BrO_3$ [M + H]⁺ 282.9964, found 282.9970.

3-(4-Methoxy-3-nitrobenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bf). Following general procedure 1 (solvent: methanol), 9b (90 mg, 0.80 mmol) and 13f (107 mg, 0.40 mmol) were converted to endo-11bf (86 mg, 0.33 mmol, 82%): yellowish liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.66 (d, J = 2.2 Hz, 1H), 7.43 (dd, J = 8.6, 2.2 Hz, 1H), 7.01 (d, J = 8.6 Hz, 1H), 6.58 (t, J = 4.3 Hz, 1H), 4.34 (t, J = 6.3 Hz, 2H), 3.91 (s, 3H), 3.58 (s, 2H), 2.50–2.41 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 164.5, 151.8, 141.0, 139.5, 135.2, 132.0, 131.1, 125.9, 113.9, 66.5, 56.7, 35.9, 24.5; IR (ATR) ν 1711 (s), 1624 (m), 1527 (s), 1351 (m), 1281 (w), 1261 (s), 1116 (s); HRMS (EI) calcd for C₁₃H₁₃NO₅ [M⁺] 263.0794, found 263.0803.

3-(4-Hydroxy-3-nitrobenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bg). Following general procedure 1 (solvent: methanol), 9b (90 mg, 0.80 mmol) and 13g (101 mg, 0.40 mmol) were converted to endo-11bg (98 mg, 0.39 mmol, 99%): yellow solid; mp 90–92 °C; ¹H NMR (600 MHz, CDCl₃) δ 10.48 (s, 1H), 7.93 (d, J = 2.1 Hz, 1H), 7.49 (dd, J = 8.5, 2.2 Hz, 1H), 7.09 (d, J = 8.6 Hz, 1H), 6.60 (t, J = 4.3 Hz, 1H), 4.37 (t, J = 6.1 Hz, 2H), 3.60 (d, J = 1.2 Hz, 2H), 2.57–2.39 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 164.4, 154.0, 141.0, 138.8, 133.5, 132.1, 131.1, 124.8, 120.2, 66.6, 36.1, 24.6; IR (ATR) ν 3281 (bw), 1715 (s), 1629 (m), 1536 (s), 1329 (m), 1116 (s); HRMS (EI) calcd for C₁₂H₁₁NO₅ [M⁺] 249.0637, found 249.0647.

3-(4-Methoxy-2-nitrobenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bh). Following general procedure 1 (solvent: methanol), 9b (90 mg, 0.80 mmol) and 13g (107 mg, 0.40 mmol) were converted to endo-11bh (51 mg, 0.19 mmol, 48%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.43 (d, J = 2.7 Hz, 1H), 7.33 (d, J = 8.6 Hz, 1H), 7.08 (dd, J = 8.6, 2.7 Hz, 1H), 6.43 (t, J = 4.2 Hz, 1H), 4.32 (t, J = 6.2 Hz, 2H), 3.86 (d, J = 1.4 Hz, 2H), 3.84 (s, 3H), 2.55–2.26 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 164.5, 158.9, 149.8, 140.7, 134.1, 130.9, 124.9, 119.8, 109.7, 66.4, 55.9, 33.5, 24.5; IR (ATR) ν 1711 (s), 1524 (s), 1505 (m), 1251 (s), 1114 (s); HRMS (EI) calcd for C₁₃H₁₃NO₅ [M⁺] 263.0794, found 263.0792.

3-(4-Nitrobenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bj). Following general procedure 1 (solvent: methanol), 9b (90 mg, 0.80 mmol) and 13j (95 mg, 0.40 mmol) were converted to endo-11bj (62 mg, 0.27 mmol, 67%): yellowish liquid; $^1{\rm H}$ NMR (300 MHz, C₆D₆) δ 7.84 (d, J=8.7 Hz, 2H), 6.82 (d, J=8.8 Hz, 2H), 5.62 (t, J=4.0 Hz, 1H), 3.60 (t, J=6.2 Hz, 2H), 3.29 (s, 2H), 1.49–1.39 (m, 2H); $^{13}{\rm C}$ NMR (75 MHz, C₆D₆) δ 163.6, 147.1, 146.5, 140.5, 131.6, 129.9, 123.7, 65.9, 37.0, 24.3; IR (ATR) ν 1709 (s), 1512 (s), 1341 (s), 1110 (s); HRMS (ESI) calcd for $\rm C_{12}H_{12}NO_4$ [M + H]+ 234.0761, found 234.0762.

3-(2-Methylcarboxylate-4-hydroxybenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bl). Following general procedure 1 (solvent: methanol), 9b (90 mg, 0.80 mmol) and 13l (107 mg, 0.40 mmol) were converted to endo-11bl (51 mg, 0.20 mmol, 49%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.43 (d, J = 2.6 Hz, 1H), 7.27 (s (br.), 1H), 7.09 (d, J = 8.3 Hz, 1H), 6.96 (dd, J = 8.3, 2.7 Hz, 1H), 6.32–6.25 (m, 1H), 4.31 (t, J = 6.2 Hz, 2H), 3.87 (s, 2H), 3.79 (s, 3H), 2.48–2.22 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 167.9, 166.0, 155.1, 140.3, 133.3, 132.8, 130.8, 130.8, 119.7, 117.8, 66.6, 52.2, 34.0, 24.4; IR (ATR) n 1714 (s), 1688 (s), 1435 (m), 1274 (s), 1216 (s); HRMS (EI) calcd for $C_{14}H_{14}O_5$ [M⁺] 262.0841, found 262.0853.

3-(4-Acetamidobenzyl)-5,6-dihydro-2H-pyran-2-one (endo-11bm). Following general procedure 2 (solvent: acetonitrile), 9b (90 mg, 0.80 mmol) and 13m (100 mg, 0.40 mmol) were converted to endo-11bm (30 mg, 0.12 mmol, 31%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 8.15 (s, 1H), 7.42 (d, J = 8.4 Hz, 2H), 7.09 (d, J = 8.3

Hz, 2H), 6.47 (t, J = 4.0 Hz, 1H), 4.32 (t, J = 6.3 Hz, 2H), 3.53 (s, 2H), 2.51–2.34 (m, 2H), 2.11 (s, 3H); 13 C NMR (75 MHz, CDCl₃) δ 168.9, 165.1, 140.6, 136.8, 134.0, 132.7, 129.6, 120.3, 66.5, 36.4, 24.5, 24.4; HRMS (EI) calcd for $C_{14}H_{15}NO_3$ [M⁺] 245.1052, found 245.1048.

3-(4-Methoxybenzyl)-6-methyl-5,6-dihydro-2H-pyran-2-one (endo-11ca). Following general procedure 2 (solvent: methanol), 9c (126 mg, 1.00 mmol) and 13a (111 mg, 0.50 mmol) were converted to endo-11ca (95 mg, 0.41 mmol, 82%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.13 (d, J=8.6 Hz, 2H), 6.85 (d, J=8.6 Hz, 2H), 6.35 (t, J=4.3 Hz, 1H), 4.58–4.44 (m, 1H), 3.79 (s, 3H), 3.57 (s, 2H), 2.33–2.25 (m, 2H), 1.41 (d, J=6.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 165.5, 158.2, 139.2, 132.8, 130.4, 130.2, 114.0, 74.2, 55.2, 35.7, 31.4, 20.7; IR (ATR) ν 1711 (s), 1511 (s), 1242 (s), 1119 (m); HRMS (ESI) calcd for C₁₄H₁₇O₃ [M + H]⁺ 233.1178, found 233.1164.

3-(4-Methoxybenzyl)dihydrofuran-2(3H)-one (16aa).⁸⁵ Arene diazonium salt 13a (89 mg, 0.40 mmol) and Pd(OAc)₂ (4.5 mg, 5.0 mol %) were suspended in methanol (4.0 mL). Lactone 9a (78 mg, 0.80 mmol) was added, and the reaction mixture was stirred at ambient temperature until the evolution of nitrogen gas had ceased. Activated charcoal (45 mg) was added, the solution was flushed with hydrogen, and the reaction mixture was kept under an atmosphere of hydrogen for 12 h. The solvent was evaporated, and the residue was partitioned between water (10 mL) and MTBE (10 mL). The aqueous layer was separated and extracted three times with MTBE (10 mL). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures of increasing polarity as eluent, to furnish 16aa (65 mg, 0.32 mmol, 79%): colorless liquid; ¹H NMR (500 MHz, CDCl₃) δ 7.13 (d, J = 8.7 Hz, 2H), 6.85 (d, J = 8.7 Hz, 2H), 4.21 (td, J = 8.8, 3.2 Hz, 1H), 4.14 (td, J = 9.2, 6.8 Hz, 1H), 3.80 (s, 3H), 3.16 (dd, J = 13.8, 4.2 Hz, 1H), 2.82 (dddd, J = 18.5, 9.5, 8.8, 4.2 Hz, 1H), 2.73 (dd, *J* = 13.8, 9.1 Hz, 1H), 2.25 (dddd, *J* = 12.6, 8.7, 6.8, 3.2 Hz, 1H), 2.04–1.94 (m, 1H); 13 C NMR (125 MHz, CDCl₃) δ 179.3, 158.8, 130.7, 130.3, 114.5, 67.0, 55.7, 41.6, 35.7, 28.3; IR (ATR) ν 1762 (s), 1512 (s), 1246 (s), 1022 (s); HRMS (ESI) calcd for $C_{12}H_{15}O_3 [M + H]^+ 207.1021$, found 207.1037.

3-(4-Hydroxybenzyl)dihydrofuren-2(3H)-one (16ab). Following the procedure given above for 16aa, lactone 9a (78 mg, 0.80 mmol) and diazonium salt 13b (83 mg, 0.40 mmol) were converted to 16ab (74 mg, 0.38 mmol, 95%): colorless liquid; ¹H NMR (500 MHz, CDCl₃) δ 7.07 (s (br.), 1H), 7.03 (d, J = 8.5 Hz, 2H), 6.82 (d, J = 8.5 Hz, 2H), 4.22–4.10 (m, 2H), 3.10 (dd, J = 13.9, 4.4 Hz, 1H), 2.83 (dddd, J = 18.4, 9.5, 8.9, 4.4 Hz, 1H), 2.72 (dd, J = 13.9, 8.8 Hz, 1H), 2.23 (dddd, J = 12.5, 8.8, 6.8, 3.6 Hz, 1H), 2.03–1.93 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 180.7, 155.4, 130.5, 130.0, 116.1, 65.7, 41.8, 35.5, 28.1; IR (ATR) ν 3357 (bm), 1742 (s), 1515 (s), 1205 (m); HRMS (ESI) calcd for C₁₁H₁₃O₃ [M + H]⁺ 193.0865, found 193.0866.

(Z)-2-(4-Methoxybenzyl)penta-2,4-dienoic Acid (Z-15ba). To a solution of endo-11ba (21.8 mg, 0.10 mmol) in DMSO (1.0 mL) was added KOH (22.4 mg, 0.40 mmol), and the reaction mixture was stirred at 45 °C until the starting material was fully consumed as indicated by TLC (ca. 1 h). After the mixture was cooled to ambient temperature, aq HCl (1 M, 10 mL) was added followed by ethyl acetate (10 mL). The aqueous layer was separated and extracted three times with ethyl acetate (10 mL for each extraction). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures as eluent, to furnish Z-15ba (15.4 mg, 0.07 mmol, 71%): colorless liquid; ¹H NMR (300 MHz, acetone- d_6) δ 7.37 (ddd, J = 17.0, 11.0, 10.2 Hz, 1H), 7.16 (d, J = 8.7, 2H), 6.86 (d, J = 8.7, 2H)8.7, 2H), 6.51 (d, J = 11.1 Hz, 1H), 5.44 (dd, J = 17.0, 1.7 Hz, 1H), 5.34 (dd, I = 10.0, 1.8 Hz, 1H), 3.77 (s, 3H), 3.61 (s, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 167.8, 158.7, 139.9, 134.5, 132.7, 131.7, 130.1, 122.7, 114.1, 54.9, 39.6; IR (ATR) ν 2941 (bw), 1676 (s), 1511 (s), 1242 (s); HRMS (EI) calcd for C₁₃H₁₄O₃ [M⁺] 218.0943, found 218.0934.

(2Z,4E)-2-(4-Methoxybenzyl)hexa-2,4-dienoic Acid ((2Z,4E)-15ca). Following the procedure given above for Z-15ba, endo-11ca (23.7 mg, 0.10 mmol) was converted to (2Z,4E)-15ca (16.4 mg, 0.07 mmol, 69%): colorless liquid; 1 H NMR (600 MHz, CDCl₃) δ 7.23 (ddq, J = 15.0, 11.3, 1.6 Hz, 1H), 7.15 (d, J = 8.7 Hz, 2H), 6.86 (d, J = 8.7 Hz, 2H), 6.50 (d, J = 11.3 Hz, 1H), 6.04 (dq, J = 15.0, 6.8 Hz, 1H), 3.82 (s, 3H), 3.60 (s, 2H), 1.89 (dd, J = 6.9, 1.3 Hz, 3H); 13 C NMR (150 MHz, CDCl₃) δ 173.1, 158.5, 144.4, 139.6, 132.0, 130.2, 129.5, 127.0, 114.2, 55.6, 39.3, 19.1; IR (ATR) ν 2929 (bw), 1677 (s), 1511 (s), 1247 (s); HRMS (EI) calcd for $C_{14}H_{17}O_3$ [M $^+$] 233.1178, found 233.1181.

General Procedure 3: Matsuda—Heck Couplings of 9d under Base-Free Conditions. The appropriate arene diazonium salt 13 (0.80 mmol) and $Pd(OAc)_2$ (4.5 mg, 5.0 mol %) were suspended in the corresponding solvent as indicated in Table 3 (4.0 mL). N-Benzyl- α -methylene- δ -valerolactam 9d (81 mg, 0.40 mmol) was added, and the reaction mixture was stirred at ambient temperature for 12 h. The solvent was evaporated, and the residue was purified by column chromatography on silica using hexane—MTBE mixtures of increasing polarity as eluent.

General Procedure 4: Matsuda—Heck Couplings of 9d under Basic Conditions. The appropriate arene diazonium salt 13 (0.80 mmol), Pd(OAc)₂ (4.5 mg, 5.0 mol %), and NaOAc (98 mg, 1.20 mmol) were suspended in the corresponding solvent as indicated in Table 3 (4.0 mL). N-Benzyl-α-methylene-δ-valerolactam 9d (81 mg, 0.40 mmol) was added, and the reaction mixture was stirred at ambient temperature for 12 h. All volatiles were evaporated, and the residue was partitioned between water (10 mL) and MTBE (10 mL). The aqueous layer was separated and extracted with MTBE (3 times, 10 mL for each extraction). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica using hexane—MTBE mixtures of increasing polarity as eluent.

1-Benzyl-3-(4-methoxybenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11da). Following the general procedure 4 (solvent: acetonitrile), 9d (81 mg, 0.40 mmol) and 13a (177 mg, 0.80 mmol) were converted to endo-11da (90 mg, 0.29 mmol, 72%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 7.41–7.19 (m, 5H), 7.15 (d, J = 8.4 Hz, 2H), 6.88 (d, J = 8.5 Hz, 2H), 6.08–6.01 (m, 1H), 4.62 (s, 2H), 3.77 (s, 3H), 3.60 (s, 2H), 3.26 (t, J = 7.1 Hz, 2H), 2.35–2.10 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 165.3, 158.0, 137.7, 135.9, 132.3, 131.7, 130.3, 128.6, 127.9, 127.3, 113.8, 55.3, 50.1, 44.8, 35.9, 23.9; IR (ATR) ν 1663 (m), 1621 (s), 1508 (s), 1242 (s), 1031 (m); HRMS (EI) calcd for $C_{20}H_{21}NO_2$ [M^+] 307.1572, found 307.1585.

1-Benzyl-3-(4-hydroxybenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11db). Following general procedure 3 (solvent: methanol), 9d (81 mg, 0.40 mmol) and 13b (166 mg, 0.80 mmol) were converted to endo-11db (101 mg, 0.34 mmol, 86%): colorless solid, mp 123–124 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.36–7.17 (m, SH), 7.04 (d, J = 8.4 Hz, 2H), 6.73 (d, J = 8.4 Hz, 2H), 6.11 (t, J = 4.3 Hz, 1H), 4.64 (s, 2H), 3.58 (s, 2H), 3.30 (t, J = 7.1 Hz, 2H), 2.33–2.19 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 165.6, 154.6, 137.6, 135.9, 134.7, 131.2, 130.4, 128.7, 128.1, 127.5, 155.5, 50.4, 45.0, 36.2, 24.4; IR (ATR) ν 3256 (bm), 1661 (m), 1607 (s), 1592 (s), 1514 (s); HRMS (EI) calcd for $C_{19}H_{19}NO_2$ [M $^+$] 293.1416, found 293.1410.

1-Benzyl-3-(4-(benzyloxy)benzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dc). Following general procedure 4 (solvent: acetonitrile), 9d (81 mg, 0.40 mmol) and 13c (238 mg, 0.80 mmol) were converted to endo-11dc (100 mg, 0.26 mmol, 65%): colorless solid, mp 189–190 °C; ¹H NMR (300 MHz, acetone- d_6) δ 7.48 (d, J = 8.4 Hz, 2H), 7.43–7.22 (m, 8H), 7.18 (d, J = 8.4 Hz, 2H), 6.93 (d, J = 8.5 Hz, 2H), 6.23 (t, J = 4.1 Hz, 1H), 5.09 (s, 2H), 4.60 (s, 2H), 3.55 (s, 2H), 2.06 (t, J = 7.1 Hz, 2H), 2.28–2.25 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 169.7, 162.5, 143.6, 142.9, 140.8, 139.7, 137.7, 135.2, 133.6, 133.6, 133.0, 132.9, 132.7, 132.2, 119.7, 74.7, 54.7, 50.0, 40.9, 29.0; IR (ATR) ν 1664 (m), 1622 (s), 1507 (s); HRMS (EI) calcd for $C_{26}H_{25}NO_2$ [M⁺] 383.1885, found 383.1882.

1-Benzyl-3-(3-bromo-4-methoxybenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dd). Following general procedure 3 (solvent: methanol), 9d (81 mg, 0.40 mmol) and 13d (240 mg, 0.80 mmol)

were converted to *endo*-**11dd** (117 mg, 0.30 mmol, 76%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 7.40 (s, 1H), 7.33–7.24 (m, 5H), 7.16 (d, J = 8.4 Hz, 1H), 6.82 (d, J = 8.3 Hz, 1H), 6.12 (t, J = 3.8 Hz, 1H), 4.62 (s, 2H), 3.86 (s, 3H), 3.59 (s, 2H), 3.28 (t, J = 7.1 Hz, 2H), 2.35–2.18 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 165.0, 154.2, 137.6, 135.3, 134.7, 133.7, 133.5, 129.4, 128.6, 127.9, 127.3, 111.9, 111.4, 56.3, 50.1, 44.8, 35.7, 23.9; IR (ATR) ν 1663 (m), 1621 (s), 1495 (s), 1252 (m), 1055 (m); HRMS (EI) calcd for $C_{20}H_{20}N^{79}BrO_{2}$ [M $^{+}$] 385.0677, found 385.0667.

1-Benzyl-3-(4-(benzyloxy)-3-bromobenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dn). Following general procedure 3 (solvent: methanol), 9d (81 mg, 0.40 mmol) and 13n (301 mg, 0.80 mmol) were converted to endo-11dn (122 mg, 0.26 mmol, 66%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.54–7.44 (m, 3H), 7.44–7.20 (m, 8H), 7.14 (dd, J = 8.3, 1.6 Hz, 1H), 6.89 (d, J = 8.4 Hz, 1H), 6.13 (t, J = 4.1 Hz, 1H), 5.14 (s, 2H), 4.64 (s, 2H), 3.60 (s, 2H), 3.29 (t, J = 7.1 Hz, 2H), 2.35–2.19 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 165.0, 153.4, 137.6, 136.7, 135.2, 134.8, 133.9, 133.8, 129.2, 128.6, 128.6, 127.9, 127.9, 127.3, 127.0, 113.9, 112.3, 70.9, 50.1, 44.8, 35.7, 23.9; IR (ATR) ν 1664 (m), 1619 (s), 1490 (s); HRMS (EI) calcd for $C_{26}H_{25}N^{79}BrO_{2}$ [M + H]⁺ 462.1063, found 462.1088.

1-Benzyl-3-(4-methoxy-3-nitrobenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11df). Following general procedure 3 (solvent: methanol), 9d (56 mg, 0.28 mmol) and 13f (150 mg, 0.56 mmol) were converted to endo-11df (79 mg, 0.22 mmol, 80%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.72 (d, J=2.0 Hz, 1H), 7.49 (dd, J=8.6, 2.0 Hz, 1H), 7.38–7.12 (m, SH), 7.03 (d, J=8.6 Hz, 1H), 6.24 (t, J=4.1 Hz, 1H), 4.62 (s, 2H), 3.93 (s, 3H), 3.67 (s, 2H), 3.31 (t, J=7.1 Hz, 2H), 2.34–2.38 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 164.7, 151.4, 139.4, 137.4, 135.4, 135.2, 134.6, 132.5, 128.6, 127.9, 127.4, 125.7, 113.6, 56.6, 50.1, 44.8, 35.9, 24.0; IR (ATR) ν 1665 (m), 1620 (s), 1528 (s); HRMS (ESI) calcd for C₂₀H₂₁N₂O₄ [M + H]⁺ 353.1496, found 353.1514.

1-Benzyl-3-(4-hydroxy-3-nitrobenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dg). Following general procedure 3 (solvent: methanol), 9d (81 mg, 0.40 mmol) and 13g (202 mg, 0.80 mmol) were converted to endo-11dg (129 mg, 0.38 mmol, 96%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 10.49 (s, 1H), 7.96 (d, J = 1.6 Hz, 1H), 7.54 (dd, J = 8.6, 1.8 Hz, 1H), 7.39–7.19 (m, 5H), 7.09 (d, J = 8.6 Hz, 1H), 6.27 (t, J = 3.9 Hz, 1H), 4.62 (s, 2H), 3.65 (s, 2H), 3.32 (t, J = 7.1 Hz, 2H), 2.43–2.21 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 164.7, 153.6, 138.9, 137.4, 135.5, 134.5, 133.4, 132.5, 128.6, 127.9, 127.4, 124.5, 119.8, 50.1, 44.8, 36.0, 23.0; IR (ATR) ν 3246 (bw), 1662 (m), 1620 (s), 1536 (s), 1342 (s); HRMS (EI) calcd for C₁₉H₁₈N₂O₄ [M⁺] 338.1267, found 338.1282.

1-Benzyl-3-(4-nitrobenzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dj). Following general procedure 3 (solvent: methanol), 9d (81 mg, 0.40 mmol) and 13j (189 mg, 0.80 mmol) were converted to endo-11dj (44 mg, 0.14 mmol, 34%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 8.15 (d, J = 8.7 Hz, 2H), 7.43 (d, J = 8.6 Hz, 2H), 7.38–7.19 (m, 5H), 6.27 (t, J = 4.0 Hz, 1H), 4.62 (s, 2H), 3.76 (s, 2H), 3.33 (t, J = 7.1 Hz, 2H), 2.36–2.30 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 164.7, 148.1, 146.6, 137.5, 135.9, 134.4, 129.9, 128.7, 128.0, 127.6, 123.7, 50.2, 44.9, 37.2, 24.1; IR (ATR) ν 1664 (m), 1621 (s), 1512 (s), 1342 (s); HRMS (EI) calcd for C₁₉H₁₈N₂O₃ [M⁺] 322.1317, found 322.1313.

1-Benzyl-3-(4-hydroxy-3-(methylbenzoate)benzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dk). Following general procedure 3 (solvent: methanol), 9d (81 mg, 0.40 mmol) and 13k (212 mg, 0.80 mmol) were converted to endo-11dk (93 mg, 0.26 mmol, 66%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 10.66 (s, 1H), 7.73 (d, J = 1.9 Hz, 1H), 7.36 (dd, J = 8.6, 2.0 Hz, 1H), 7.49—7.16 (m, 5H), 6.93 (d, J = 8.5 Hz, 1H), 6.12 (t, J = 4.0 Hz, 1H), 4.63 (s, 2H), 3.93 (s, 3H), 3.63 (s, 2H), 3.30 (t, J = 7.1 Hz, 2H), 2.31—2.25 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 170.6, 165.1, 160.1, 137.6, 136.9, 135.5, 134.6, 130.4, 130.2, 128.6, 128.0, 127.5, 117.4, 112.1, 52.3, 50.1, 44.8, 36.0, 24.0; IR (ATR) ν 3190 (bw), 1664 (s). 1620 (s), 1439 (m), 1209 (s), 1090 (m); HRMS (ESI) calcd. for $C_{21}H_{22}NO_4$ [M + H] $^+$ 352.1543, found 352.1563.

1-Benzyl-3-(4-hydroxy-2-(methylbenzoate)benzyl)-5,6-dihydropyridin-2-(1H)-one (endo-11dl). Following general procedure 3 (solvent: methanol), 9d (66 mg, 0.34 mmol) and 13l (181 mg, 0.68 mmol) were converted to endo-11dl (77 mg, 0.22 mmol, 65%): colorless liquid; 1 H NMR (300 MHz, CDCl₃) δ 7.99 (s, 1H), 7.44 (d, J=2.5 Hz, 1H), 7.37–7.14 (m, 5H), 7.08 (d, J=8.4 Hz, 1H), 6.93 (dd, J=8.3, 2.5 Hz, 1H), 5.94 (t, J=4.3 Hz, 1H), 4.68 (s, 2H), 3.95 (s, 2H), 3.75 (s, 3H), 3.30 (t, J=7.1 Hz, 2H), 2.24–2.20 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 168.0, 165.9, 155.1, 137.3, 135.5, 134.7, 132.9, 131.7, 130.8, 128.7, 128.0, 127.5, 119.6, 117.7, 52.0, 50.4, 44.9, 34.1, 23.9; IR (ATR) ν 3207 (bw), 1716 (m), 1597 (s), 1293 (m). 7-Hydroxychroman-2one (18). 86 A suspension of 17 (3.24 g, 20.0

7-Hydroxychroman-2one (*18*). ⁸⁶ A suspension of 17 (3.24 g, 20.0 mmol) and Pd/C (200 mg, 10 wt %) in glacial acetic acid (60 mL) was stirred under an atmosphere of hydrogen for 12 h. The solvent was evaporated, and the residue purified by column chromatography on silica using hexane/MTBE mixtures of increasing polarity as eluent to furnish 18 (3.24 g, 19.7 mmol, 99%): colorless solid, mp 134–136 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.04 (d, J = 8.2 Hz, 1H), 6.66–6.58 (m, 2H), 5.74 (bs, 1H), 2.96–2.89 (m, 2H), 2.82–2.72 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 169.3, 156.0, 152.7, 128.8, 114.5, 111.9, 104.5, 29.6, 23.1; IR (ATR) ν 3340 (bw), 1764 (m), 1627 (m), 1141 (s), 1106 (s); HRMS (EI) calcd for C₉H₈O₃ [M⁺] 164.0473, found 164.0475.

7-((tert-Butyldimethylsilyl)oxy)chroman-2-one (19).87 A solution of 18 (3.00 g, 18.3 mmol) in CH₂Cl₂ (50 mL) was cooled to 0 °C, and NEt₃ (2.80 mL, 27.5 mmol) was added, followed by dropwise addition of a solution of TBSCl (4.10 g, 27.5 mmol) in CH₂Cl₂ (20 mL). After addition was completed, the reaction mixture was warmed to ambient temperature and stirred until complete conversion of the starting material. The reaction was quenched by addition of water (50 mL), the layers were separated, and the aqueous layer was extracted three times with CH2Cl2 (25 mL for each extraction). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures of increasing polarity as eluent, to furnish 19 (4.63 g, 19.7 mmol, 91%): colorless liquid; ¹H NMR (300 MHz, CDCl₃) δ 7.01 (d, J = 8.1 Hz, 1H), 6.57 (dd, J = 8.1, 2.4 Hz, 1H), 6.54 (m, 2H), 2.96–2.88 (m, 2H), 2.81–2.68 (m, 2H), 0.97 (s, 9H), 0.19 (s, 6H); 13 C NMR (75 MHz, CDCl₃) δ 168.6, 155.8, 152.6, 128.4, 116.3, 115.3, 108.8, 29.6, 25.7, 23.2, 18.3, -4.4; IR (ATR) ν 2930 (w), 1767 (s), 1620 (m), 1504 (s), 1134 (s), 1105 (s); HRMS (EI) calcd for C₁₅H₂₂O₃Si [M⁺] 278.1338, found 278.1339.

7-((tert-Butyldimethylsilyl)oxy)-3-methylenchroman-2-one (20). To a solution of diisopropylamine (2.53 mL, 18.0 mmol) in THF (50 mL) was added BuLi (2.5 M solution in hexane, 7.20 mL, 18.0 mmol) dropwise at -78 °C. After 0.25 h, a solution of 19 (1.67 g, 6.0 mmol) in THF (10 mL) was added dropwise, and stirring was continued for 1 h at -78 °C, followed by addition of Eschenmoser's salt (3.88 g, 21.0 mmol) in one portion. The reaction mixture was then warmed to ambient temperature and stirred for 12 h. A satured aqueous solution of NH₄Cl (25 mL) was added, and the layers were separated. The aqueous layer was extracted three times with diethyl ether (25 mL). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was dissolved in THF (20 mL), and methyl iodide (1.87 mL, 30.0 mmol) was added. The reaction mixutere was stirred for 12 h at ambient temperature. The suspension was filtered through a pad of Celite and washed with diethyl ether (25 mL). All volatiles were evaporated, and the residue was purified by column chromatography on silica, using hexane/ MTBE mixtures of increasing polarity as eluent, to furnish 20 (0.97 g, 3.4 mmol, 56%): colorless solid; mp 46-47 °C; ¹H NMR (300 MHz, C_6D_6) δ 6.63 (d, J = 2.1 Hz, 1H), 6.58 (d, J = 8.1 Hz, 1H), 6.52 (dd, J= 8.2, 2.3 Hz, 1H), 6.17-6.14 (m, 1H), 5.04-5.00 (m, 1H), 2.98 (s, 2H), 0.94 (s, 9H), 0.07 (s, 6H); 13 C NMR (75 MHz, C_6D_6) δ 162.3, 156.0, 152.1, 132.5, 128.4, 127.4, 116.5, 114.6, 109.0, 31.3, 25.8, 18.4, - 4.5; IR (ATR) ν 2930 (w), 2858 (w), 1751 (m), 1622 (m), 1504 (s), 1151 (s), 1099 (s); HRMS (EI) calcd for $C_{16}H_{22}O_3Si\ [M^+]$ 290.1338, found 290.1346.

7-Hydroxy-3-methylenchroman-2-one (21). To a solution of 20 (100 mg, 0.34 mmol) in THF (10 mL) was added a solution of TBAF-

 $3H_2O$ (108 mg, 0.34 mmol) in THF (3 mL) at -78 °C. After full conversion of the starting material, the reaction mixture was hydrolyzed with a saturated aqueous solution of NH₄Cl (3 mL) and warmed to ambient temperature. Diethyl ether (15 mL) was added, and the layers were separated. The aqueous layer was extracted three times with diethyl ether (15 mL for each extraction). The combined organic extracts were dried with MgSO₄, filtered, and evaporated. The residue was purified by column chromatography on silica, using hexane/MTBE mixtures of increasing polarity as eluent, to furnish 21 (55 mg, 0.31 mmol, 91%): colorless solid, decomposition at 230 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.01 (d, J = 8.2 Hz, 1H), 6.70 (d, J =2.2 Hz, 1H), 6.63 (dd, J = 8.2, 2.3 Hz, 1H), 6.42 (s, 1H), 5.81 (bs, 1H), 5.78 (s, 1H), 3.73 (s, 2H); 13 C NMR (75 MHz, CDCl₃) δ 163.8, 155.9, 151.3, 131.9, 128.9, 128.5, 112.9, 112.1, 104.4, 31.2; IR (ATR) ν 3363 (bm), 1723 (s), 1628 (s), 1298 (s), 1153 (s); HRMS (EI) calcd for C₁₀H₈O₃ [M⁺] 176.0473, found 176.0478.

Anemarcoumarin A. To a suspension of phenoldiazonium salt 13b (31 mg, 0.15 mmol) and Pd(OAc)₂ (1.7 mg, 5.0 mol %) in methanol (3.0 mL) was added 21 (53 mg, 0.30 mmol), and the reaction mixture was stirred at ambient temperature until the gas evolution had ceased. The solvent was evaporated, and the residue was purified by column chromatography on silica, using hexane/MTBE mixtures of increasing polarity as eluent, to furnish anemarcoumarin A (24.5 mg, 0.09 mmol, 61%): colorless solid; mp 206–209 °C; ¹H NMR (600 MHz, methanol- d_4) δ 7.33 (s, 1H), 7.22 (d, J = 8.5 Hz, 1H), 7.04 (d, J = 8.5 Hz, 2H), 6.70 (d, J = 8.5 Hz, 2H), 6.68 (dd, J = 8.5, 2.3 Hz, 1H) 6.61 (d, J = 2.3 Hz, 1H), 3.61 (s, 2H); 13 C NMR (150 MHz, methanol- d_4) δ 164.2, 162.0, 157.1, 156.0, 141.5, 131.2, 130.4, 130.0, 126.0, 116.3, 114.3, 113.6, 103.0, 36.4; IR (ATR) ν 3324 (bm), 1688 (m), 1612 (s), 1513 (m), 1233 (m); HRMS (EI) calcd for $C_{16}H_{13}O_4$ [M + H]* 269.0808, found 269.0822. All analytical data match those reported for the natural product. ⁶⁶

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b02207.

¹H and ¹³C NMR spectra for all compounds (PDF) Documentation of the DFT calculations (tables of atom coordinates and absolute energies) (PDF)

AUTHOR INFORMATION

Corresponding Author

*E-mail: bernd.schmidt@uni-potsdam.de.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We thank Evonik Oxeno for generous donations of solvents and Umicore (Hanau, Germany) for generous donations of catalysts.

REFERENCES

- (1) The Mizoroki-Heck Reaction; Oestreich, M., Ed.; Wiley: Chichester, 2009.
- (2) Cabri, W.; Candiani, I. Acc. Chem. Res. 1995, 28, 2-7.
- (3) Jutand, A. In *The Mizoroki-Heck Reaction*; Oestreich, M., Ed.; Wiley: Chichester, 2009; pp 1-50.
- (4) Lautens, M.; Fang, Y.-Q. Org. Lett. 2003, 5, 3679-3682.
- (5) Kamisaki, H.; Nanjo, T.; Tsukano, C.; Takemoto, Y. Chem. Eur. J. 2011, 17, 626–633.
- (6) Canterbury, D. P.; Hesp, K. D.; Polivkova, J. Org. Biomol. Chem. 2016, 14, 7731–7734.
- (7) Kirschbaum, S.; Waldmann, H. Tetrahedron Lett. **1997**, 38, 2829–
- (8) Tanaka, D.; Myers, A. G. Org. Lett. 2004, 6, 433-436.

- (9) Brunner, H.; de Courcy, N. L. C.; Genet, J. P. Tetrahedron Lett. 1999, 40, 4815-4818.
- (10) Arcadi, A.; Chiarini, M.; Marinelli, F.; Berente, Z.; Kollàr, L. Org. Lett. 2000, 2, 69-72.
- (11) Arcadi, A.; Chiarini, M.; Marinelli, F.; Berente, Z.; Kollár, L. Eur. J. Org. Chem. 2001, 3165-3173.
- (12) Kim, S. H.; Kim, K. H.; Lee, H. J.; Kim, J. N. Tetrahedron Lett. 2013, 54, 329-334.
- (13) Ding, Y.-H.; Fan, H.-X.; Long, J.; Zhang, Q.; Chen, Y. Bioorg. Med. Chem. Lett. 2013, 23, 6087-6092.
- (14) Schmidt, B.; Hölter, F.; Berger, R.; Jessel, S. Adv. Synth. Catal. 2010, 352, 2463-2473.
- (15) Schmidt, B.; Hölter, F.; Kelling, A.; Schilde, U. J. Org. Chem. 2011, 76, 3357-3365.
- (16) Schmidt, B.; Elizarov, N. Chem. Commun. 2012, 48, 4350-4352.
- (17) Schmidt, B.; Elizarov, N.; Berger, R.; Holter, F. Org. Biomol. Chem. 2013, 11, 3674-3691.
- (18) Stern, T.; Rückbrod, S.; Czekelius, C.; Donner, C.; Brunner, H. Adv. Synth. Catal. 2010, 352, 1983-1992.
- (19) Felpin, F.-X.; Coste, J.; Zakri, C.; Fouquet, E. Chem. Eur. J. 2009, 15, 7238-7245.
- (20) Barancelli, D. A.; Salles, A. G.; Taylor, J. G.; Correia, C. R. D. Org. Lett. 2012, 14, 6036-6039.
- (21) Selvakumar, K.; Zapf, A.; Spannenberg, A.; Beller, M. Chem. -Eur. J. 2002, 8, 3901-3906.
- (22) Raduán, M.; Padrosa, J.; Pla-Quintana, A.; Parella, T.; Roglans, A. Adv. Synth. Catal. 2011, 353, 2003-2012.
- (23) Li, Z.; Ip, F. C. F.; Ip, N. Y.; Tong, R. Chem. Eur. J. 2015, 21, 11152-11157.
- (24) Kikukawa, K.; Matsuda, T. Chem. Lett. 1977, 6, 159-162.
- (25) Kikukawa, K.; Nagira, K.; Wada, F.; Matsuda, K. Tetrahedron 1981, 37, 31-36.
- (26) Felpin, F.-X.; Nassar-Hardy, L.; Le Callonnec, F.; Fouquet, E. Tetrahedron 2011, 67, 2815-2831.
- (27) Roglans, A.; Pla-Quintana, A.; Moreno-Mañas, M. Chem. Rev. 2006, 106, 4622-4643.
- (28) Taylor, J. G.; Moro, A. V.; Correia, C. R. D. Eur. J. Org. Chem. 2011, 1403-1428.
- (29) Schmidt, B.; Berger, R.; Hölter, F. Org. Biomol. Chem. 2010, 8,
- (30) Susperregui, N.; Miqueu, K.; Sotiropoulos, J.-M.; Le Callonnec, F.; Fouquet, E.; Felpin, F.-X. Chem. - Eur. J. 2012, 18, 7210-7218.
- (31) Oger, N.; Le Grognec, E.; Felpin, F.-X. J. Org. Chem. 2014, 79, 8255-8262.
- (32) Schmidt, B.; Wolf, F.; Brunner, H. Eur. J. Org. Chem. 2016, 2972-2982.
- (33) Felpin, F.-X.; Miqueu, K.; Sotiropoulos, J.-M.; Fouquet, E.; Ibarguren, O.; Laudien, J. Chem. - Eur. J. 2010, 16, 5191-5204.
- (34) Tanaka, A.; Yamashita, K. Agric. Biol. Chem. 1978, 42, 1585-1588.
- (35) Murray, A. W.; Reid, R. G. Synthesis 1985, 35-38.
- (36) Song, L.; Yao, H.; Zhu, L.; Tong, R. Org. Lett. 2013, 15, 6-9.
- (37) Krawczyk, H.; Śliwiński, M. Tetrahedron 2003, 59, 9199-9211.
- (38) Ichimoto, I.; Machiya, K.; Kirihata, M.; Ueda, H. Agric. Biol. Chem. 1990, 54, 657-662.
- (39) Jacobs, W. A.; Craig, L. C. J. Am. Chem. Soc. 1938, 60, 1701-1702.
- (40) Lee, D. L.; Morrow, C. J.; Rapoport, H. J. Org. Chem. 1974, 39, 893-902.
- (41) Schmidt, B.; Elizarov, N.; Schilde, U.; Kelling, A. J. Org. Chem. 2015, 80, 4223-4234.
- (42) Erb, W.; Hellal, A.; Albini, M.; Rouden, J.; Blanchet, J. Chem. -Eur. J. 2014, 20, 6608-6612.
- (43) Salabert, J.; Sebastián, R. M.; Vallribera, A.; Cívicos, J. F.; Nájera, C. Tetrahedron 2013, 69, 2655-2659.
- (44) Gaikwad, D. S.; Pore, D. M. Synlett 2012, 23, 2631-2634.
- (45) Kutonova, K. V.; Trusova, M. E.; Stankevich, A. V.; Postnikov, P. S.; Filimonov, V. D. Beilstein J. Org. Chem. 2015, 11, 358-362.

- (46) Schmidt, B.; Elizarov, N.; Riemer, N.; Hölter, F. Eur. J. Org. Chem. 2015, 5826-5841.
- (47) Machado, A. H. L.; Milagre, H. M. S.; Eberlin, L. S.; Sabino, A. A.; Correia, C. R. D.; Eberlin, M. N. Org. Biomol. Chem. 2013, 11, 3277-3281.
- (48) Severino, E. A.; Costenaro, E. R.; Garcia, A. L. L.; Correia, C. R. D. Org. Lett. 2003, 5, 305-308.
- (49) Hofmann, A. W. Annalen 1859, 110, 129-140.
- (50) Cardillo, G.; Orena, M.; Sandri, S. Tetrahedron 1976, 32, 107-
- (51) Corey, E. J.; Schmidt, G. Tetrahedron Lett. 1979, 20, 2317-2320.
- (52) Roush, W. R.; Spada, A. P. Tetrahedron Lett. 1982, 23, 3773-3776.
- (53) Constantino, M. G.; Losco, P.; Castellano, E. E. J. Org. Chem. 1989, 54, 681-683.
- (54) Schmidt, B.; Kunz, O. Eur. J. Org. Chem. 2012, 1008-1018.
- (55) Schmidt, B.; Kunz, O. Org. Lett. 2013, 15, 4470-4473.
- (56) Schmidt, B.; Audörsch, S. Org. Lett. 2016, 18, 1162-1165.
- (57) Savanur, H. M.; Kalkhambkar, R. G.; Laali, K. K. Tetrahedron Lett. 2016, 57, 663-667.
- (58) Machotta, A. B.; Oestreich, M. In The Mizoroki-Heck Reaction; Oestreich, M., Ed.; Wiley: Chichester, 2009; pp 179-213.
- (59) Larock, R. C.; Gong, W. H.; Baker, B. E. Tetrahedron Lett. 1989, 30, 2603-2606.
- (60) Grigg, R.; Loganathan, V.; Santhakumar, V.; Sridharan, V.; Teasdale, A. Tetrahedron Lett. 1991, 32, 687-690.
- (61) Sabino, A. S.; Machado, A. H. L.; Correia, C. R. D.; Eberlin, M. N. Angew. Chem., Int. Ed. 2004, 43, 2514-2518.
- (62) Schmidt, B. Chem. Commun. 2003, 1656-1657.
- (63) Awale, S.; Miyamoto, T.; Linn, T. Z.; Li, F.; Win, N. N.; Tezuka, Y.; Esumi, H.; Kadota, S. J. Nat. Prod. 2009, 72, 1631-1636.
- (64) Li, M.-C.; Yao, Z.; Takaishi, Y.; Tang, S.-A.; Duan, H.-Q. Chem. Biodiversity 2011, 8, 1112-1120.
- (65) Feng, S.; Xu, L.; Wu, M.; Hao, J.; Qiu, S. X.; Wei, X. Fitoterapia 2010, 81, 472-474.
- (66) Youn, U. J.; Lee, Y. S.; Jeong, H.; Lee, J.; Nam, J.-W.; Lee, Y. J.; Hwang, E. S.; Lee, J.-H.; Lee, D.; Kang, S. S.; Seo, E.-K. J. Nat. Prod. 2009, 72, 1895-1898.
- (67) Rempel, V.; Volz, N.; Hinz, S.; Karcz, T.; Meliciani, I.; Nieger, M.; Wenzel, W.; Bräse, S.; Müller, C. E. J. Med. Chem. 2012, 55, 7967-
- (68) Rempel, V.; Volz, N.; Gläser, F.; Nieger, M.; Bräse, S.; Müller, C. E. J. Med. Chem. 2013, 56, 4798-4810.
- (69) Behrenswerth, A.; Volz, N.; Toräng, J.; Hinz, S.; Bräse, S.; Müller, C. E. Bioorg. Med. Chem. 2009, 17, 2842-2851.
- (70) Cheng, J.-F.; Chen, M.; Wallace, D.; Tith, S.; Arrhenius, T.; Kashiwagi, H.; Ono, Y.; Ishikawa, A.; Sato, H.; Kozono, T.; Sato, H.; Nadzan, A. M. Bioorg. Med. Chem. Lett. 2004, 14, 2411-2415.
- (71) Dube, P. N.; Waghmare, M. N.; Mokale, S. N. Chem. Biol. Drug Des. 2016, 87, 608-617.
- (72) Toräng, J.; Vanderheiden, S.; Nieger, M.; Bräse, S. Eur. J. Org. Chem. 2007, 2007, 943-952.
- (73) Schreiber, J.; Maag, H.; Hashimoto, N.; Eschenmoser, A. Angew. Chem., Int. Ed. Engl. 1971, 10, 330-331.
- (74) Paquette, L. A.; Backhaus, D.; Braun, R. J. Am. Chem. Soc. 1996, 118, 11990-11991.
- (75) Lee, C.; Yang, W.; Parr, R. G. Phys. Rev. B: Condens. Matter Mater. Phys. 1988, 37, 785-789.
- (76) Becke, A. D. J. Chem. Phys. 1993, 98, 5648-5652.
- (77) Hertwig, R. H.; Koch, W. Chem. Phys. Lett. 1997, 268, 345–351.
- (78) Neese, F. WIREs Comput. Mol. Sci. 2012, 2, 73-78.
- (79) Weigend, F.; Ahlrichs, R. Phys. Chem. Chem. Phys. 2005, 7, 3297-3305.
- (80) Andrae, D.; Häußermann, U.; Dolg, M.; Stoll, H.; Preuß, H. Theor. Chim. Acta 1990, 77, 123-141.
- (81) Sinnecker, S.; Rajendran, A.; Klamt, A.; Diedenhofen, M.; Neese, F. J. Phys. Chem. A 2006, 110, 2235-2245.

- (82) Fu, M.; Chen, L.; Jiang, Y.; Jiang, Z.-X.; Yang, Z. Org. Lett. 2016, 18, 348–351.
- (83) Li, S.; Ma, S. Chem. Asian J. 2012, 7, 2411-2418.
- (84) Tanaka, M.; Mitsuhashi, H.; Maruno, M.; Wakamatsu, T. Chem. Lett. 1994, 23, 1455–1458.
- (85) Ito, M.; Ootsuka, T.; Watari, R.; Shiibashi, A.; Himizu, A.; Ikariya, T. J. Am. Chem. Soc. 2011, 133, 4240–4242.
- (86) Hernández-Galán, R.; Salvá, J.; Massanet, G. M.; Collado, I. G. Tetrahedron 1993, 49, 1701–1710.
- (87) Kamat, V. S.; Graden, D. W.; Lynn, D. G.; Steffens, J. C.; Riopel, J. L. Tetrahedron Lett. 1982, 23, 1541–1544.